# New Fossils of the Oldest North American Euprimate Teilhardina brandti (Omomyidae) from the Paleocene-Eocene Thermal Maximum 

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#### Abstract

More than 25 new specimens of Teilhardina brandti, one of the oldest known euprimates, are reported from earliest Eocene strata of the southern Bighorn Basin, Wyoming. The new fossils include the first upper dentitions, a dentary showing the lower dental formula for the first time, and the first postcrania ascribed to T. brandti (tarsals and terminal phalanges). The elongated navicular and long talar neck suggest that $T$. brandti was an active arboreal quadruped, and the terminal phalanges constitute the oldest evidence for nails in


Euprimates. Phylogenetic analysis incorporating the new data indicates that T. brandti is more derived than $T$. belgica but less so than T. americana. The hypothesis that Teilhardina originated in Asia (T. asiatica) and dispersed westward to Europe (T. belgica) and then to North America (T. brandti and T. magnoliana) during the earliest Eocene Paleocene-Eocene Thermal Maximum is most consistent with available evidence, including the relative age of fossil samples and their stage of evolution. Am J Phys Anthropol 000:000-000, 2011. © 2011 Wiley-Liss, Inc.

Except for the poorly known and enigmatic Altiatlasius from the late Paleocene of Morocco, euprimates are unknown in the fossil record until the beginning of the Eocene, when they abruptly appear, almost simultaneously, in Europe, Asia, and North America (Ni et al., 2004; Smith et al., 2006). In North America the omomyid Teilhardina brandti is arguably the oldest known euprimate, recorded from basal Eocene (earliest Wasatchian, Wa-0) sediments of northern Wyoming deposited during the Paleocene-Eocene Thermal Maximum or PETM (Gingerich, 1993; Smith et al., 2006). Carbon isotope stratigraphy indicates that T. brandti first appeared in the Bighorn Basin, Wyoming, $\sim 25$ Kyr after the onset of the global carbon isotope excursion (CIE), which was coincident with the PETM. Based on dental differences among T. asiatica, T. belgica, and T. brandti, and their stratigraphic positions relative to the minimum value of the CIE, Smith et al. (2006) hypothesized that Teilhardina dispersed from eastern Asia to Europe and from there to North America during the first 25 Kyr of the PETM. In contrast, Beard (2008) argued that Teilhardina magnoliana, from the uppermost Tuscahoma Formation Red Hot local fauna of Mississippi, predated both T. brandti from the Bighorn Basin and T. belgica from the Tienen Formation at Dormaal, Belgium. He further hypothesized that Teilhardina first reached coastal areas of North America from Asia during the beginning of the PETM, later dispersing to Europe and the Bighorn Basin. Current evidence suggests that the adapoid primate Cantius did not appear in the southern Bighorn Basin until somewhat later in the PETM (Rose et al., in press), which makes Teilhardina the oldest North American euprimate. Dispersal of primates and other mammals was probably facilitated by the opening of high-latitude land connections between the northern continents
associated with global warming during the PETM (McKenna, 1983; Krause and Maas, 1990; Clyde and Gingerich, 1998).

Teilhardina brandti was initially based on a single lower molar (Gingerich, 1993), and until now the species has been known only from a small number of fragmentary lower dentitions (Smith et al., 2006). Here we report 29 new specimens of $T$. brandti, including the first known upper dentitions, a lower jaw that shows for the first time the size and number of lower antemolar teeth, and the first postcrania attributed to this species, which include the oldest definitive evidence for the presence of nails in euprimates. In addition, a cladistic analysis of Teilhardina species, taking into account new anatomical information from these specimens, allows for re-evaluation of previously published, and highly debated, phylogenetic and biogeographic hypotheses of the first Euprimates to appear in North America, Europe, and Asia.

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## GEOLOGICAL SETTING AND AGE

Teilhardina brandti is restricted to the second earliest biozone of the early Eocene, Wa-0, which immediately follows the brief Wa-M biozone and coincides with most of the PETM. Besides including the oldest North American euprimates, the Wa-0 fauna is characterized by the first appearance of perissodactyls, artiodactyls, and hyaenodontid creodonts (Gingerich, 1989; Koch et al., 1992), and by small body size of some herbivorous and carnivorous mammals (Gingerich, 1989; Chester et al., 2010). The PETM was an episode of global warming, $\sim 150 \mathrm{Kyr}$ in duration (Aubry et al., 2007; Abdul Aziz et al., 2008), first detected from the CIE in marine sediments (e.g., Kennett and Stott, 1991; Zachos et al., 2001). Subsequently the PETM has also been recognized in continental sections on several continents based on carbon and oxygen isotope measurements from soil carbonates, hematite coatings on fossil bones, and fossil teeth (e.g., Koch et al., 1992, 2003; Fricke et al., 1998; Bao et al., 1999). The onset of the CIE is now used to mark the beginning of the Eocene, at $\sim 55.8 \mathrm{Ma}$ (Aubry et al., 2007).

The fossils reported here come from Wa-0 strata of the Willwood Formation, in the Sand Creek Divide and Cabin Fork sections of the southern Bighorn Basin, Wyoming. The primate fossils come chiefly from paleosols and small channel cuts into paleosols. Carbon isotope stratigraphy in the Sand Creek Divide and Cabin Fork sections places the lowest occurrence of Teilhardina

|  | Abbreviations |
| :---: | :---: |
| AMNH | Department of Vertebrate Paleontology, American Museum of Natural History, New York, New York |
| CM | Section of Vertebrate Paleontology, Carnegie Museum of Natural History, Pittsburgh, Pennsylvania |
| GU | Department of Geology, H.N.B. Garhwal University, Srinagar, Uttarakhand, India |
| IRSNB | Institut Royal des Sciences Naturelles de Belgique, Brussels, Belgium |
| IVPP | Institute of Vertebrate Paleontology and Paleoanthropology, Beijing, China |
| LACM | Los Angeles County Museum of Natural History, Los Angeles, California |
| MNH | Muséum National d'Histoire Naturelle, Paris, France |
| PSS | Geological Institute Paleontology and Stratigraphy Section, Mongolian Academy of Sciences, Ulan Bator, Mongolian People's Republic |
| RS | Richard Smith collection, Wemmel, Belgium |
| UCMP | University of California Museum of Paleontology, Berkeley, California |
| UF | Vertebrate Paleontology Collection, Florida Museum of Natural History, University of Florida, Gainesville, Florida |
| UKMNH | University of Kansas Museum of Natural History, Lawrence, Kansas |
| UM | University of Michigan Museum of Paleontology, Ann Arbor, Michigan |
| USGS | US Geological Survey, Denver, Colorado; collection transferred to USNM |
| USNM | Department of Paleobiology, National Museum of Natural History, Smithsonian Institution, Washington, D.C. |
| UW | Department of Geology and Geophysics, University of Wyoming, Laramie, Wyoming |
| YPM | Department of Vertebrate Paleontology, Peabody Museum of Natural History, Yale University, New Haven, Connecticut. |

brandti between 8 and 10 m above the onset of the CIE (see Fig. 1) and $\sim 5-10 \mathrm{~m}$ above the minimum $\delta^{13} \mathrm{C}$ values (Rose et al., in press). This corresponds to the approximate date of the first appearance of T. brandti in the Polecat Bench section (northern Bighorn Basin), at $\sim 25 \mathrm{Kyr}$ after the start of the CIE and $\sim 10 \mathrm{Kyr}$ after the minimum $\delta^{13} \mathrm{C}$ values (Bains et al., 2003; Magioncalda et al., 2004; Smith et al., 2006).

## MATERIALS AND METHODS

We report here 23 new dental specimens (Tables 1 and 2) of Teilhardina brandti and attribute several isolated postcranial elements to this species. Specimens were recovered through both surface prospecting and screen-washing.

## Morphology

Comparisons were made with other species of Teilhardina as well as a diversity of other primitive euprimates and plesiadapiforms. Lower tooth characters used in the phylogenetic analysis were primarily taken or modified from those of Smith et al. (2006) and Beard (2008). These are the most recent phylogenetic analyses focusing on Teilhardina, but both studies employed only lower tooth characters (upper teeth were previously unknown for T. brandti). Therefore, upper tooth characters were based mainly on those used by Seiffert et al. (2005), whose analysis encompassed a broad array of fossil primates including Teilhardina and a diversity of other omomyids. Some characters used in these previous analyses were omitted because they were deemed uninformative, ambiguous, or unrepeatable. Lengths and widths are maximum dimensions unless otherwise noted.

Particular attention was paid to characters of $\mathrm{P}_{4}$ that have been considered to be informative for Teilhardina (see Fig. 2)-including tooth width, metaconid height (Bown and Rose, 1987; Smith et al., 2006; Beard, 2008), preprotocristid verticality (Beard, 2008), and abruptness of the transition from the postvallid to the buccal surface of the trigonid (Beard, 2008)—with the intention of quantifying previous qualitative observations. Trigonid length of $\mathrm{P}_{4}$ was measured as the maximum distance from the mesial margin to the postvallid surface directly posterior to the protoconid, with the tooth viewed occlusally. To facilitate consistent measurements, we generated 3D digital models of $36 \mathrm{P}_{4}$ s representing Teilhardina species, other early euprimates, and plesiadapiforms (see Appendix D). Models were generated by scanning casts of isolated teeth using a Scanco $\mu$ CT 40, with settings of $70 \mathrm{kv}, 114 \mu \mathrm{Amp}$, and voxel resolution of $8 \mu \mathrm{~m}$. A LDI RPS-120 laser scanner was used to generate models of $\mathrm{P}_{4}$ for the adapoids Marcgodinotius and Asiadapis. Resulting scans were processed and measured in Avizo 6.0 (see Boyer, 2008, for details on processing of microCT data). Digital photographs were measured for several comparative taxa (Steinius, Tetonius, Cantius, Donrussellia, Altanius, Purgatorius, and Palaechthon). The measurements and variables we present (see Appendix $D$ and $E$ ) are those that best capture the originally described variation. Resulting values were incorporated into the phylogenetic analysis by assigning character states to different ranges of values (Appendix E).

In addition, we used principal coordinates analysis and the statistics software PAST.exe to analyze the significance of the various $\mathrm{P}_{4}$ measurements noted above


Fig. 1. Stratigraphic occurrences of Teilhardina brandti from the southern Bighorn Basin, Wyoming. Fossils are from two different stratigraphic sections, the bases of which are correlated via the onset of the Carbon Isotope Excursion (CIE), documented in dispersed organic carbon of the two sections (Wing et al., 2005; Rose et al., in press). The bold dashed line at the top is correlated at the base of a prominent red bed (Big Red) that contains the stratigraphically highest paleosol nodules (PSN). Close similarity in stratigraphic thickness (meters) suggests relative uniformity in depositional rates across the $10-20$ miles separating the two regions. Possible correlative beds are shown by dotted lines. Blue arrows indicate localities that yielded T. brandti. Sand Creek Divide localities (with prefix WW) were established by Johns Hopkins University, Cabin Fork localities (with prefix WY) by the University of Florida. Sand Creek Divide section by Mary J. Kraus. Colors of beds roughly match appearance of paleosols in the field; yellow are immature, red and purple more mature. Sands are represented by speckled units. T. brandti first appears about $8-10 \mathrm{~m}$ after the onset of the CIE. Other abbreviations: BR-1, Big Red paleosol 1; CS-1, Carbonaceous Shale 1; LR, Little Red paleosol; (S)LIRB, (Second) Lowest Intermittent Red Bed; P-x, Purple-x; RGR - Red-Grey-Red; Str-Gr - strange gray.

TABLE 1. Measurements of upper teeth of Teilhardina brandti and T. americana ${ }^{\text {a }}$

|  | Locality | $\mathrm{P}^{3} \mathrm{~L}$ | $\mathrm{P}^{3} \mathrm{~W}$ | $\mathrm{P}^{4} \mathrm{~L}$ | $\mathrm{P}^{4} \mathrm{~W}$ | $\mathrm{M}^{1} \mathrm{~L}$ | $\mathrm{M}^{1} \mathrm{~W}$ | $\mathrm{M}^{2} \mathrm{~L}$ | $\mathrm{M}^{2} \mathrm{~W}$ | $\mathrm{M}^{3} \mathrm{~L}$ | $\mathrm{M}^{3} \mathrm{~W}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Teilhardina brandti |  |  |  |  |  |  |  |  |  |  |  |
| USNM 540598 | WW-74 |  |  |  |  |  |  | 1.80 | 3.10 |  |  |
| UF 244453 | WY0017 |  |  |  |  |  |  | 1.70 | 2.90 |  |  |
| UF 244460 | WY0014 |  |  |  |  |  |  |  |  | 1.15 | 2.20 |
| UF 254927 | WY0025 |  |  |  |  | 1.8* | 2.80 | 1.65 | 3.20 |  |  |
| USNM 525543 | WW-84 | 1.55 | 2.00 |  |  |  |  |  |  |  |  |
| USNM 533494 | WW-74 |  |  | 1.60 | 2.40 | 1.75 | 2.70 | 1.70 | 2.90 | 1.10 | 2.05 |
| USNM 539466 | WW-74 |  |  |  |  |  |  | 1.70 | 2.85 | 1.15 | 2.10 |
| USNM 539467 | WW-74 |  |  | 1.60 | 2.35 | 1.75 | 2.70 | 1.80 | 3.00 | 1.20 | 2.00 |
| $N$ |  | 1 | 1 | 2 | 2 | 3 | 3 | 6 | 6 | 4 | 4 |
| Mean |  |  |  |  | 2.38 | 1.77 | 2.73 | 1.73 | 2.99 | 1.15 | 2.09 |
| Standard deviation |  |  |  |  | 0.04 | 0.03 | 0.06 | 0.06 | 0.14 | 0.04 | 0.09 |
| Standard error |  |  |  |  | 0.03 | 0.02 | 0.03 | 0.03 | 0.06 | 0.02 | 0.04 |
| Minimum |  |  |  |  | 2.35 | 1.75 | 2.70 | 1.65 | 2.85 | 1.10 | 2.00 |
| Maximum |  |  |  |  | 2.40 | 1.80 | 2.80 | 1.80 | 3.20 | 1.20 | 2.20 |
| Teilhardina americana <br> UF 244459 | WY0016 | 1.70 | 2.10 | 1.65 | 2.30 | 1.80 | 2.70 | 1.85 | 3.25 | 1.25 | 2.40 |

${ }^{\mathrm{a}} \mathrm{L}$ of upper molars $=$ maximum L parallel to a line through paracone-metacone; $\mathrm{W}=$ maximum distance perpendicular to L .
*Denotes approximate. All specimens are new.
TABLE 2. Measurements of lower teeth of Teilhardina brandti

|  | Locality | $\mathrm{P}_{4} \mathrm{~L}$ | $\mathrm{P}_{4} \mathrm{~W}$ | $\mathrm{M}_{1} \mathrm{~L}$ | $\mathrm{M}_{1}$ Wtri | $\mathrm{M}_{1}$ Wtal | $\mathrm{M}_{2} \mathrm{~L}$ | $\mathrm{M}_{2}$ Wtri | $\mathrm{M}_{2}$ Wtal | $\mathrm{M}_{3} \mathrm{~L}$ | $\mathrm{M}_{3} \mathrm{~W}$ tri | $\mathrm{M}_{3} \mathrm{~W}$ tal | $\mathrm{M}_{3} \mathrm{~W}$ hyd |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| UF 244451 | WY0015 |  |  |  |  |  | 2.05 | 1.75 | 1.75 | 2.10 | 1.40 | 1.15 | 0.55 |
| UF 244454 | WY0019 | 1.80 | 1.35 |  |  |  |  |  |  |  |  |  |  |
| UF 244455 | WY0020 |  |  | 2.10 |  | 1.60 | 2.00 | 1.60 | 1.55 |  |  |  |  |
| UF 244456 | WY0021 |  |  |  |  |  | 2.00 | 1.60 | 1.60 | 1.90 | 1.30 | 1.10 | 0.45 |
| UF 244457 | WY0022 |  |  |  |  |  | 1.85 | 1.50 | 1.40 |  |  |  |  |
| UF 254928 | WY0020 |  |  |  |  |  |  |  |  | 1.95 | 1.40 | 1.10 | 0.60 |
| UF 254929 | WY0027 |  |  | 1.90 | 1.30 | 1.45 | 1.95 | 1.65 | 1.50 |  |  |  |  |
| UF 254931 | WY0026 |  |  | 1.90 | 1.30 | 1.45 |  |  |  |  |  |  |  |
| UF 254935 | WY0029 |  |  |  |  |  |  |  |  | 1.95 | 1.35 | 1.10 | 0.60 |
| UM 99031 HT | SC-351 |  |  |  |  |  | 1.95 | 1.55 | 1.50 |  |  |  |  |
| UM 111434 | SC-67 |  | 1.30 | 2.05 | 1.40 | 1.50 | 2.15 | 1.70 | 1.60 | 1.90 | 1.20 | 0.90 |  |
| USNM 493913 | WW-108 | 1.60 | 1.20 | 1.95 | 1.40 | 1.60 |  |  |  |  |  |  |  |
| USNM 493914 | WW-107 |  |  |  |  |  | 1.95 | 1.60 | 1.60 | 1.95 | 1.30 | 1.05 |  |
| USNM 521795 | WW-75 |  |  |  |  |  | 2.00 | 1.55 | 1.60 |  |  |  |  |
| USNM 525543 | WW-84 | 1.75 | 1.30 | 1.95 | 1.30 | 1.45 | 1.95 | 1.60 | 1.55 |  |  |  |  |
| USNM 525544 | WW-84 | 1.75 | 1.20 |  |  |  | 2.00 | 1.55 | 1.55 |  |  |  |  |
| USNM 525545 | WW-97 |  |  |  |  |  | 1.95 | 1.50 | 1.50 |  |  |  |  |
| USNM 525546 | WW-96 |  |  |  |  |  | 1.90 | 1.50 | 1.50 | 1.90 | 1.35 | 1.15 |  |
| USNM 525621 | WW-125 |  |  | 2.00 | 1.40 | 1.55 | 2.05 | 1.70 | 1.60 | 2.10 | 1.30 | 1.05 |  |
| USNM 525622 | WW-96 |  |  |  |  |  | 1.95 | 1.45 | 1.45 | 1.80 | 1.25 | 1.05 |  |
| USNM 533505 | WW-74 |  |  |  |  |  | 2.00 | 1.50 | 1.45 |  |  |  |  |
| USNM 533554 | WW-77 | 1.80 | 1.20 | 2.05 | 1.40 | 1.50 |  |  |  |  |  |  |  |
| USNM 538082L | WW-84 |  |  | 2.00 | 1.25 | 1.35 | 2.00 | 1.55 | 1.50 |  |  |  |  |
| USNM 538082R | WW-84 |  |  |  |  |  | 1.95 | 1.60 | 1.50 |  |  |  |  |
| USNM 538084 | WW-84 |  |  |  |  |  | 1.95 | 1.50 | 1.45 |  |  |  |  |
| $N$ |  | 5 | 6 | 9 | 8 | 9 | 19 | 19 | 19 | 9 | 9 | 9 | 4 |
| Mean |  | 1.74 | 1.26 | 1.99 | 1.34 | 1.49 | 1.98 | 1.58 | 1.53 | 1.95 | 1.32 | 1.07 | 0.55 |
| Standard deviation |  | 0.08 | 0.07 | 0.06 | 0.06 | 0.08 | 0.06 | 0.08 | 0.08 | 0.10 | 0.07 | 0.08 | 0.07 |
| Standard error |  | 0.04 | 0.03 | 0.02 | 0.02 | 0.03 | 0.01 | 0.02 | 0.02 | 0.03 | 0.02 | 0.03 | 0.04 |
| Minimum |  | 1.60 | 1.20 | 1.90 | 1.25 | 1.35 | 1.85 | 1.45 | 1.40 | 1.80 | 1.20 | 0.90 | 0.45 |
| Maximum |  | 1.80 | 1.35 | 2.10 | 1.40 | 1.60 | 2.15 | 1.75 | 1.75 | 2.10 | 1.40 | 1.15 | 0.60 |

Specimens in bold are new.
Abbreviations: L, length; W, width; W hyd, width of hypoconulid lobe; Wtal, width of talonid; Wtri, width of trigonid.
(buccolingual angle [character \#26], preprotocristid verticality angle [\#29], trigonid width index [\#31], and mesial angle [\#46]). Angles were analyzed in radians, thus having a magnitude and range of values similar to the $\mathrm{P}_{4}$ trigonid width index. Metaconid height was excluded because this cusp is absent (Purgatorius) or variable (Marcgodinotius and Asiadapis) in some taxa in the analysis; however, results of an analysis including metaconid measurements on a reduced sample did not differ
substantially from those excluding the measurements. The $\mathrm{P}_{4}$ analysis included two euprimate outgroups (Purgatorius, Palaechthon), the probable basal euprimate Altanius orlovi, and several primitive euprimates (Donrussellia gallica, Marcgodinotius indicus, Asiadapis cambayensis, Cantius ralstoni) as well as six species of Teilhardina and two other omomyids (Tetonius and Steinius) (see Appendix D). A minimum spanning tree was imposed on the principal coordinate morphospace to


Fig. 2. Measurements of $P_{4}$. Abbreviations: B, buccal; L, lingual; Ms, mesial; Oc, occlusal; preprot, preprotocristid. Scale $=$ 1 mm . See text and Appendix B for measurement definitions.
show nearest neighbors reflecting the four-dimensional distances between specimens.

## Phylogenetic analysis

To assess relationships among Teilhardina species, a matrix including 13 taxa and 47 dental characters (see Appendices A, B) was assembled in MacClade 4.06 (Maddison and Maddison, 2003). All characters and taxa were reevaluated for this study. Our analysis included six species of Teilhardina: T. asiatica, T. belgica, T. brandti, T. magnoliana, T. americana, and T. crassidens. Other basal euprimates included in the analysis were the omomyids Tetonius matthewi and Steinius vespertinus, and the primitive adapoids Donrussellia provincialis and D. gallica. The probable basal euprimate Altanius orlovi, and the basal plesiadapiforms Purgatorius janisae and Palaechthon nacimienti, were also included. A branch and bound search was conducted in PAUP* 4.0b10 for Macintosh PPC (Swofford, 2003). Purgatorius janisae was set as the outgroup. All characters were treated as unordered and were weighted equally. Multistate characters were interpreted as polymorphic.

## Descriptive paleontology

Family OMOMYIDAE Trouessart, 1879
Teilhardina Simpson, 1940
Teilhardina brandti Gingerich, 1993
New dental specimens-USNM 533494 ( R maxilla, $\mathrm{P}^{4}$ $\mathrm{M}^{3}$ ), 533505 (R dentary, $\mathrm{M}_{2}$ ), 533554 (L dentary, $\mathrm{P}_{4}-\mathrm{M}_{1}$ ), 538082 (isolated teeth: $\mathrm{LM}_{1}, \mathrm{LM}_{2}, \mathrm{RM}_{2}$ ), $538084\left(\mathrm{LM}_{2}\right)$, 538361 ( L dentary, $\mathrm{M}_{2}$ talonid), 539466 ( L maxilla, $\mathrm{M}^{2-3}$ ), 539467 (L maxilla, $\mathrm{P}^{4}-\mathrm{M}^{3}$ ), $540598\left(\mathrm{RM}^{2}\right)$; UF 244451 ( R dentary, $\mathrm{M}_{2-3}$ ), $244453\left(\mathrm{RM}^{2}\right), 244454$ ( R dentary, $\mathrm{P}_{4}$ ), 244455 ( L dentary, $\mathrm{M}_{1-2}$ ), 244456 ( L dentary, $\mathrm{M}_{2-3}$ ), 244457 ( R dentary, $\mathrm{M}_{2}$ ), 254927 ( R maxilla, $\mathrm{M}^{1-2}$ ), 254928 ( L dentary, $\mathrm{M}_{3}$ ), 254929 ( R dentary, $\mathrm{M}_{1-2}$ and all anterior alveoli), $254931\left(\mathrm{RM}_{1}\right), 254935$ ( R dentary, $\mathrm{M}_{3}$ ), and questionably $244460\left(\mathrm{LM}^{2-3}\right)$.

Maxilla and upper teeth. The maxilla is very shallow below the orbit, and the lingual root of $\mathrm{M}^{2}$ protrudes into the orbital floor. The lower rim of the orbit is present in each of the more complete maxillae (see Fig. 3), but it appears to be too incomplete to provide a confident estimate of orbital diameter.

Alveoli in USNM 533494 (Fig. 3A,B) indicate that $\mathrm{P}^{3}$ was a three-rooted triangular tooth, about as long mesio-
distally as $\mathrm{P}^{4}$ but less transverse buccolingually, as in other species of Teilhardina and closely allied taxa. $\mathrm{P}^{4}$ is markedly transverse, slightly waisted, and shorter mesiodistally than the molars. It is dominated by the large and high buccal cusp (paracone), the tallest cusp of the postcanine toothrow, which tilts distally. Although the base of the cusp is centrally positioned, the apex is distinctly distal to the buccolingual midline axis. A tiny parastyle is present but there is no discernible metastyle. The protocone is low and situated near the mesiolingual margin of the tooth. A weak postprotocrista descends distally to the postcingulum; a stronger preprotocrista runs buccally and becomes continuous with the precingulum, which extends to the parastyle.
The upper molars (Fig. 3A-C) are among the most primitive known for euprimates. The enamel is smooth, with little or no evidence of crenulation. The molar pattern is simple, with small conules, weak styles, no hypocone, and moderately developed cingula (buccally, mesially, and distally, but not lingually). A tiny parastyle is evident on all three molars, but the metastyle is indistinct. The postcingulum extends lingually slightly farther than the precingulum, but both end near the base of the protocone. At its lingual extent, the postcingulum is slightly broader than the precingulum, but there is no hypocone, and the cingulum is broadly discontinuous lingually. A faint swelling can be detected at the distolingual angle of the postcingulum, where a small hypocone may develop in later species. There is no trace of a nannopithex fold except in $\mathrm{M}^{2}$ of USNM 539467, in which a faint elevation of the enamel in the appropriate location can be detected with reflected light; however, it is clearly not a typical nannopithex fold.

UF 244460 (Fig. 3F) differs from other upper molars of T. brandti in having weakly crenulated enamel, a nearly complete lingual cingulum, and a squared lingual margin of $\mathrm{M}^{2}$ associated with a weak nannopithex fold. All of these features are known to vary in Teilhardina americana (which comes from the Wa-1/2 biozone) but are typically present in that species. Although this might suggest that UF 244460 actually comes from above Wa-0, local stratigraphy makes such contamination very unlikely. Thus it is probable that this specimen indicates intraspecific variation in T. brandti and provides evidence for a close (probably ancestor-descendant) relationship between T. brandti and T. americana.

Dentary and lower teeth. The dentary of Teilhardina brandti (UF 254929, Fig. 4) has an anterior mental foramen below $\mathrm{P}_{1}$ and a posterior foramen below the anterior root of $\mathrm{P}_{3}$. The symphysis extends back only to below $\mathrm{P}_{2}$.


Fig. 3. Maxillary dentitions of Teilhardina brandti and T. americana. A and B, T. brandti, USNM 533494, right maxilla with $\mathrm{P}^{4}-\mathrm{M}^{3}$ in buccal (A) and occlusal (B) views. C, T. brandti, USNM 539467, left maxilla with $\mathrm{P}^{4}-\mathrm{M}^{3}$ in occlusal view. D and E, T. americana, UF 244459, right maxilla with $\mathrm{P}^{3}-\mathrm{M}^{3}$ in buccal (D) and occlusal (E) views. F, T. brandti, UF 244460, left maxilla with $\mathrm{M}^{2-3}$ in occlusal view. Note crenulation, lingual cingulum, and nannopithex fold in E, slightly less developed in F. lc, lingual cingulum; nf, nannopithex fold; or, orbital rim.

UF 254929 (see Fig. 4) is the first specimen of T. brandti to reveal the lower dental formula (2.1.4.3) and the size and position of anterior teeth. It clearly shows the presence of four premolars, including single-rooted $P_{2}$ and $P_{1}$, the latter diminutive and displaced laterally. Judging from its alveolus, the canine was comparatively smaller than in adapoids but relatively large for an omomyid. The incisor alveoli indicate that $I_{1}$ was slightly larger than $I_{2}$ and that both were very small, much smaller than the canine, and neither alveolus being larger than that of $\mathrm{P}_{1} . \mathrm{I}_{1}$ is situated medial to $I_{2}$, rather than largely anterior to it as is the case in the primitive adapoid Cantius (Rose et al., 1999). Because they are incomplete anteriorly, the orientation of the roots is difficult to determine with certainty, but they appear to have been essentially vertical and not nearly so anteriorly inclined as in Cantius.
$\mathrm{P}_{4}$ has been considered the most important lower tooth for analyzing dental evolution in Teilhardina (Rose and Bown, 1984; Bown and Rose, 1987), so its morphology in T. brandti is of particular interest. USNM 533554 (Fig. $5 \mathrm{~A}-\mathrm{C}$ ) contains the fifth known $\mathrm{P}_{4}$ of $T$. brandti and only the second one preserved in sequence with $M_{1}$. All five $P_{4}$ specimens are relatively narrow and have a metaconid approximately two-thirds as high as the protoconid, which is essentially the same as in T. belgica and lower than in T. americana (Table 3 and Appendix D and E). The talonid is variable, ranging from very short and weakly developed (USNM 493913; Fig. 5D-F) to somewhat longer and basined (but open lingually) with a distinct cristid obliqua, hypoconid, and lower entoconid (USNM 533554).

The lower molars do not differ in any significant way from those described and illustrated by Smith et al.


Fig. 4. Lower dentition of Teilhardina brandti, UF 254929, right dentary with $\mathrm{M}_{1-2}$ and alveoli or roots of $\mathrm{I}_{1}-\mathrm{P}_{4}$, in occlusal (A) and buccal (B) views. View in A is slightly oblique, with mesial end tilted upward to show incisor alveoli more clearly. Note presence of small, nearly vertical incisors, moderately large canine, and a small, buccally displaced $\mathrm{P}_{1}$.
(2006), and the new $\mathrm{M}_{2}$ S are essentially identical to the holotype. Lower molars of Teilhardina brandti are characterized by low entoconids, but are variable with regard to expression of buccal cingulids and hypoconulids. They are comparable in length to those of T. americana and are longer than molars of T. belgica. They are relatively narrower, on average, than molars of T. americana, and similar in relative width to those of T. belgica (Fig. 6; see also Fig. 14). Body mass estimates based on $\mathrm{M}_{1}$ area ( 83.5 g using prosimian regression, 88 g using all primates) are comparable to those generated for T. americana (Conroy, 1987).

## Dental comparisons

Based mainly on the new fossils, we compared Teilhardina brandti with the most primitive species of the genus (T. asiatica, T. belgica, T. magnoliana, and T. americana), as well as with Baataromomys ulaanus and the basal adapoids Cantius and Donrussellia. These comparisons are summarized here.
Teilhardina asiatica. The maxillary thickness between the alveolar border and the orbital rim is markedly shallower in T. brandti than in T. asiatica, which might suggest larger orbits in T. brandti. Nevertheless, orbital size in T. brandti, to the extent that it can be estimated from the short preserved portion of the orbital rim, appears to be comparable to that in T. asiatica.

Ni et al. (2004) considered the dental morphology of T. asiatica to be very similar to that of T. belgica, but they described $\mathrm{M}^{1}$ of T. asiatica as having a more smoothly curved mesiolingual margin and a more "protuberant" distolingual aspect. These features could be interpreted as derived relative to T. belgica (when compared to other Teilhardina). In other respects, however, T. asiatica is plesiomorphic relative to most or all other species of Teilhardina. T. asiatica implicitly lacks a nannopithex fold (as T. americana was said to differ in
having it) and has a weaker buccal cingulum than T. americana (Ni et al., 2004). The anterior lower dentition of T. asiatica is less compressed than in all other Teilhardina, with short diastemata separating the premolars, and a $P_{1}$ that is not laterally displaced. The metaconid of $\mathrm{P}_{4}$ is very low relative to the protoconid. Compared to T. brandti, the lower molars of T. asiatica are relatively narrower (see Fig. 6) and lower crowned, and both the lower molars and $\mathrm{P}_{4}$ have slightly more acute cusps. As in T. brandti and T. belgica, the lower molar entoconids are low, and as in some T. brandti the paraconid is distinctly lower than the metaconid.

Teilhardina magnoliana. This species was recently named based on a sample of isolated teeth from Mississippi (Beard, 2008; Beard and Dawson, 2009), which are roughly $10 \%$ smaller in linear dimensions than those of T. brandti. T. magnoliana is said to differ from other species of Teilhardina except T. asiatica in $\mathrm{P}_{4}$ morphology and in having lower-crowned molars, and from T. brandti and T. americana in having relatively narrower molars (Beard, 2008). Only three $\mathrm{P}_{4} \mathrm{~S}$ are known for T. magnoliana, two of which are damaged and now lack the talonid (CM 70439 and 73229); the other one is more complete but abraded (CM 70434). Nevertheless, as the most important tooth for assessing evolutionary stage in Teilhardina, they merit attention. CM 73229 is slightly smaller than but otherwise very similar to $\mathrm{P}_{4} \mathrm{~S}$ of T. brandti and T. belgica, whereas CM 70434 differs in having a relatively wider trigonid, distinctly higher metaconid, and a more inclined (rather than nearly vertical) preprotocrista (Fig. 7, Table 3). These features contrast with those of $\mathrm{P}_{4}$ in T. asiatica and compare more favorably with the more derived T. americana than with T. brandti or T. belgica. CM 70434 also has a basined talonid, as in one specimen of T. brandti (USNM 533554, Fig. 5). Beard (2008) and Beard and Dawson (2009) did not discuss these differences among the $\mathrm{P}_{4} \mathrm{~s}$ attributed to


Fig. 5. Dentaries of Teilhardina brandti showing variation in $\mathrm{P}_{4}$. A-C, USNM 533554, left dentary with $\mathrm{P}_{4}-\mathrm{M}_{1}$, in occlusal (A), buccal (B), and lingual (C) views. D-F, USNM 493913, right dentary with $\mathrm{P}_{4}-\mathrm{M}_{1}$, same views. Note larger talonid and metaconid on $\mathrm{P}_{4}$ and stronger buccal cingulid on $\mathrm{P}_{4}-\mathrm{M}_{1}$ in USNM 533554.
T. magnoliana. If all three premolars represent T. magnoliana, it implies a substantial amount of morphological variation in this tooth in the Tuscahoma species.
T. magnoliana resembles T. belgica in its small size (although the holotype $\mathrm{M}_{2}$ is smaller than in any other species), relatively weak buccal cingulids, and relatively narrow lower molars (Fig. 6; see also Fig. 14). However, their relative narrowness is proportionate to their smaller length, so the molars of T. magnoliana are on average only slightly narrower than those of T. brandti $\left(\mathrm{M}_{1} \mathrm{~W} / \mathrm{L}=0.70\right.$ in T. magnoliana compared to 0.75 in T. brandti; $\mathrm{M}_{2} \mathrm{~W} / \mathrm{L}=$ 0.76 in T. magnoliana compared to 0.80 in T. brandti). Although the lower molars of T. magnoliana resemble those of T. asiatica in being lower crowned, on average, than both T. belgica and T. brandti, the latter two species are variable in this regard, with some specimens comparable in brachydonty to T. magnoliana.

The upper teeth of T. magnoliana are very similar to those of T. brandti except for being smaller. The few differences observed are quite subtle. $\mathrm{P}^{4}$ (CM 77210) is vir-
tually identical to that of T. brandti in expression of cingula, parastyle, and orientation and relative size of the two main cusps. However, the $\mathrm{P}^{4}$ paracone of T. magnoliana appears to be taller, and the distal aspect of the protocone is slightly more complex than in T. brandti, with a weak postprotocrista and a more lingual crest descending nearly to the base of the protocone; the two crests frame a slightly concave surface on the distal face of the protocone. Both CM 77210 and CM 70433 (which is damaged buccally) have slightly lower pre- and postcingula, barely above the base of the crown. CM 70433 lacks evidence of a postprotocrista.
$\mathrm{M}^{1-2}$ of Teilhardina magnoliana are represented by nine isolated teeth ( $\mathrm{M}^{3}$ is not known). $\mathrm{M}^{1}$ is about $10 \%$ narrower buccolingually than that of T. brandti but otherwise almost identical. It has small conules, a tiny parastyle, and an incipient metastyle. The cingula are similarly developed, but the postcingulum is slightly stronger and extends slightly farther lingually and mesially in T. magnoliana. The lingual cingulum is
variable but never strongly developed. Viewed from the lingual side, a nearly complete faint lingual cingulum is discernible in CM 67858 and especially CM 77211, but the cingulum is clearly discontinuous lingually in CM

TABLE 3. $P_{4}$ metaconid height index in Teilhardina

|  | Metaconid <br> height <br> index |  |  |  |
| :--- | :--- | :---: | :---: | :---: |
| Species | Spec. no. | Mean | Range |  |
| T. asiatica | IVPP 7165 | 0.63 |  |  |
| T. magnoliana | CM 70434 | 0.74 | 0.72 | $0.70-0.74$ |
| T. belgica | CM 73229 | IRSNB CL192 | 0.70 | 0.65 |
|  | IRSNB CL246 | 0.66 |  | $0.62-0.73$ |
|  | IRSNB CL457 | 0.66 |  |  |
|  | IRSNB M64 | 0.65 |  |  |
|  | IRSNB M4090 | 0.68 |  |  |
|  | IRSNB M4291 | 0.66 |  |  |
|  | IRSNB M4292 | 0.64 |  |  |
|  | IRSNB WL13 | 0.66 |  |  |
|  | IRSNB WL159 | 0.62 |  |  |
|  | IRSNB WL1052 | 0.69 |  |  |
|  | IRSNB WL1062 | 0.73 |  |  |
| T. brandti | IRSNB WL1180 | 0.71 | 0.69 | $0.60-0.72$ |
|  | UF 244454 | 0.69 | 0.68 |  |
|  | USNM 493913 | 0.60 |  |  |
|  | USNM 525543 | 0.72 |  |  |
|  | USNM 525544 | 0.70 |  |  |
| T. americana | USNM 533554 | 0.72 | 0.75 | $0.71-0.81$ |
|  | UF 244452 | 0.75 | 0.75 |  |
|  | UM 65770 75160 | 0.71 | 0.78 |  |
|  | UM 76600 | 0.73 |  |  |
|  | USGS 3849 | 0.81 |  |  |
|  | USGS 15406 | 0.76 |  |  |
|  | USGS 25324 | 0.72 |  |  |
|  | USNM 539489 | 0.74 |  |  |

${ }^{\text {a }}$ Metaconid height index $=$ metaconid height $1 /$ protoconid height (character 28).

70422 and CM 70431. Although a nannopithex fold is typically absent in both T. magnoliana and T. brandti, a weak, incipient fold is present in CM $70431\left(\mathrm{M}^{1}\right)$, as in $\mathrm{M}^{2}$ of T. brandti, USNM 539467. Compared to T. brandti, $\mathrm{M}^{2}$ of T. magnoliana is about $10 \%$ narrower and mesiodistally shorter lingually (the protocone is subtly less inflated) but similar in buccal length. It is otherwise identical to that of T. brandti. The three known $\mathrm{M}^{2} \mathrm{~s}$ lack both a nannopithex fold and a lingual cingulum.

These comparisons suggest that the upper teeth are virtually identical in the two species and differ in such subtle characters that they probably would not be distinguished if found together. The slightly smaller size of $T$. magnoliana might be related to its lower-latitude occurrence and possibly even warmer conditions along the Gulf Coast than in the continental interior during the early Eocene.

Teilhardina belgica. Smith et al. (2006) considered Teilhardina brandti to be very similar to T. belgica (Figs. 8A-C and 9A) and distinguished the two only by a few subtle features. T. brandti is slightly larger than T. belgica $(<10 \%)$, the crowns of the cheek teeth are slightly more basally inflated (resulting in, on average, slightly greater relative width; see Figs. 6 and 7), and the buccal cingulids are variably stronger. All of these features are derived relative to T. belgica. Although T. brandti has cheek teeth of comparable length to those of the slightly younger T. americana (also from the Willwood Formation of the Bighorn Basin), but narrower, it is plesiomorphic and more like T. belgica in having a lower metaconid on $\mathrm{P}_{4}$ (Table 3, Fig. 7), a narrower hypoconulid lobe on $\mathrm{M}_{3}$, and a lower entoconid on $\mathrm{M}_{1-2}$. For these reasons Smith et al. (2006) concluded that T. brandti is phylogenetically intermediate between T. belgica and T. americana. The larger sample now available for T. brandti shows variation in several traits


Fig. 6. Lower molar dimensions of Teilhardina species. Polygons delimit the observed range of each species (range of T. brandti is shaded). Where points appear clustered, all data points represent the same value but are spread apart to illustrate the number of points and species that overlap. Holotypes are indicated by a diamond within the point (lectotype for T. belgica). Specimens were measured to 0.05 mm except for Baataromomys (nearest 0.01 mm ; from Ni et al., 2007). T. brandti is similar in molar length to T. americana, but its talonid width is intermediate between T. belgica and T. americana.





Fig. 7. Box plots for six shape variables of $P_{4}$ in Teilhardina and other primitive euprimates. Variable numbers correspond to character list in Appendix B. Single specimen positions are indicated by black circles. Two specimens for a taxon sample are indicated by two horizontal lines connected by a vertical line. For larger samples the observed range is shown by a solid line, while shaded boxes encompass $50 \%$ of the data points and a horizontal line indicates the median. Units for angles are degrees. Abbreviations: Tas, Teilhardina asiatica; Tmg-a, T. magnoliana CM 73229; Tmg-b, T. magnoliana CM 70434; Tbg, T. belgica; Tbr, T. brandti; Ta, T. americana; Tc, T. crassidens; Stv, Steinius vespertinus; Ttm, Tetonius matthewi; Ao, Altanius orlovi; Dg, Donrussellia gallica; Cr, Cantius ralstoni; Mi, Marcgodinotius indicus; Ac, Asiadapis cambayensis. See Methods for description of measurements, and Table 3 and Appendix D for variable values and specimen numbers.
(e.g., strength of hypoconulids and buccal cingulids, relative hypsodonty, crown width) that support this phylogenetic position. In certain other traits, variation has been observed in T. belgica (Gingerich, 1977) that overlaps the condition in T. brandti. For example, $\mathrm{P}_{1}$ in T. belgica is small and laterally placed in the lectotype (IRSNB M64) and in IRSNB CL188, as in T. brandti (UF 254929), but it is somewhat larger and more centrally located in IRSNB M4296, and apparently absent in RS DIII467. The lower canine in T. belgica ranges from much larger than that of T. brandti (in the lectotype) to being comparable in size to that of T. brandti in IRSNB M4296 and RS DIII467. Judging from partial alveoli, the lectotype of T. belgica (like T. brandti) appears to have had small incisors (Gingerich, 1977), but their relative size and orientation are ambiguous.

Upper teeth of T. brandti (except for UF 244460) are virtually identical to those of T. belgica, except for being slightly larger. In the two available specimens of upper teeth of T. belgica (IRSNB WL 1299 and WL 1398) the precingulum extends slightly farther lingually than in T. brandti, but no other differences were observed.

Teilhardina americana. The lower canine in the holotype of T. americana (Fig. 8D-F) has a root of similar size to that of T. brandti; lower incisors of T. americana are unknown. $\mathrm{P}_{1}$ is variably present in T. americana (see Bown and Rose, 1987); when present it is consistently very small and laterally displaced. The lower cheek teeth of Teilhardina americana are similar in length to those of T. brandti but are slightly wider (see Fig. 6). T. americana further differs from T. brandti in having a higher metaconid on $\mathrm{P}_{4}$ (Table 3, Fig. 7), taller molar entoconids, and typically stronger cingulids and weaker hypoconulids. $\mathrm{M}_{3}$ tends to be less reduced and to have a larger hypoconulid lobe. Upper molars of T. americana are derived relative to T. brandti in having a weak nannopithex fold and, often, a small hypocone (Figs. 3D,E, 9B). A lingual cingulum is present though variably developed, and the enamel tends to be weakly crenulated. These derived features relative to T. brandti, together with presence of T. brandti in immediately underlying strata, support direct descent of $T$. americana from T. brandti.

Teilhardina crassidens occurs in strata above those that yield T. americana and has been interpreted as a


Fig. 8. Lower dentition of (A-C) Teilhardina belgica, lectotype, IRSNB M64, left $\mathrm{P}_{3}-\mathrm{M}_{3}$ and anterior alveoli, and (D-F) T. americana, holotype, UW 6896, left $\mathrm{C}_{1}-\mathrm{M}_{3}$. D-F from Bown and Rose (1987). Note that T. americana has relatively broader cheek teeth and a higher metaconid on $\mathrm{P}_{4}$ compared to T. belgica.
direct descendant of T. americana (Bown and Rose, 1987). T. crassidens is more derived than T. americana in consistently lacking $P_{1}$ and having relatively wider and more inflated cheek teeth (see Fig. 14) and a more molarized $\mathrm{P}_{4}$ with a higher metaconid, features contrasting with T. brandti. The younger species T. tenuicula and T. demissa remain rare and poorly understood. They differ from T. brandti in having relatively broad, lowcrowned $\mathrm{P}_{3-4}$, with mesiodistally extended trigonids and higher metaconids (Bown and Rose, 1987; Rose, 1995).

Baataromomys ulaanus. Ni et al. (2007) described this new omomyid based on an isolated $\mathrm{M}_{2}$ from the Bumbanian (early Eocene) of Inner Mongolia, China. They showed that $\mathrm{M}_{2} \mathrm{~S}$ of B. ulaanus and Teilhardina brandti (which they assigned to Baataromomys) are intermediate in size and proportions between Old World T. asiatica and T. belgica, on the one hand, and other North American Teilhardina on the other (see Fig. 6 herein). Baataromomys was said to differ from Teilhardina (including North American species other than T. brandti) "in having a smaller paraconid that is more closely appressed to the metaconid, and a relatively lower hypoconid and cristid obliqua," and to differ further from North American Teilhardina "in having a lower entoconid, a much weaker crest connecting the entoconid with the metaconid, and a weaker buccal cingulid" (Ni et al., 2007: p 3).

Based on our analysis, when intraspecific variation is taken into account, none of these features seems to separate Baataromomys from Teilhardina, nor do they support a special relationship between B. ulaanus and T. brandti. Comparison of multiple specimens of T. belgica, T. brandti, T. magnoliana, and T. americana indicate that molar paraconid height and position relative to the metaconid are variable in T. brandti, with most specimens closely approximating T. belgica and T. asiatica in paraconid expression. Our comparisons revealed no significant differences in height of the hypoconid or cristid
obliqua, although T. americana and T. crassidens tend to be relatively slightly higher crowned. We agree that B. ulaanus and, to a greater extent, T. brandti, differ from T. americana and T. crassidens in having a lower entoconid, but this feature is shared with T. belgica and T. asiatica and may be plesiomorphic for Teilhardina. The metaconid and entoconid are not truly connected by a crest because the talonid notch intervenes; but to the extent that crests can be compared on the lingual side of $\mathrm{M}_{2}$, again we found no significant differences between T. brandti and T. belgica. Development of the buccal cingulids is variable in Teilhardina, particularly so in T. brandti, with some specimens having weak or absent cingula as in T. belgica and others having moderate or strong cingula as in T. americana and T. crassidens (see Fig. 5). These comparisons support retention of T. brandti in Teilhardina, and call into question the generic distinction of Baataromomys.

Primitive adapoids. Compared to Wasatchian Cantius, the oldest and most primitive North American adapoid, Teilhardina brandti already exhibits the omomyid hallmark of mesiodistal compression of the dentition. Although its primitive euprimate dental formula is the same as that of Cantius (2.1.4.3), the antemolar dentition is conspicuously compacted relative to that of Cantius. The small incisors of T. brandti appear to have been more vertically implanted, with $\mathrm{I}_{1}$ slightly larger than and medial to $\mathrm{I}_{2}$. The lower canine is relatively smaller than that of Cantius. The premolars of T. brandti are especially crowded together, with both $\mathrm{P}_{1}$ and $P_{2}$ single-rooted, $P_{1}$ reduced and displaced laterally, and no diastemata between any adjacent antemolar teeth. In contrast, Cantius typically has mesiodistally extended premolars, short diastemata between anterior premolars, $\mathrm{P}_{1}$ not laterally displaced, and a two-rooted $\mathrm{P}_{2}$ (except in Wa-0 C. torresi, in which the root is bilobed). In Cantius abditus $\mathrm{I}_{1}$ is smaller than $\mathrm{I}_{2}$ and the


Fig. 9. Upper dentition of (A) Teilhardina belgica, IRSNB WL 1398, right $\mathrm{P}^{3}, \mathrm{M}^{1-3}$, and (B) T. americana, UW 8871, right $\mathrm{P}^{4}-\mathrm{M}^{3}\left(\mathrm{M}^{3}\right.$ restored from UW 8961). B from Bown and Rose (1987). lc, lingual cingulum; nf, nannopithex fold. T. americana is larger than T. belgica and further differs in having crenulated enamel, a nannopithex fold, and a variably developed lingual cingulum.
incisors are anteriorly inclined at about $45^{\circ}$ (Rose et al., 1999).

European Donrussellia, usually considered the most primitive adapoid, had a two-rooted, unreduced $\mathrm{P}_{2}$ (Godinot, 1978). $\mathrm{P}_{2}$ was also two-rooted in the asiadapine Marcgodinotius, the most primitive Asian adapoid (Rose et al., 2009). Both Donrussellia and Marcgodinotius have uncompressed antemolar dentitions and relatively larger lower canines than in T. brandti. These observations suggest that an uncompressed antemolar dentition with a relatively large canine and unreduced, two-rooted $\mathrm{P}_{2}$ is primitive for adapoids. The metaconid on $\mathrm{P}_{4}$ is very low in Donrussellia and asiadapines (when present), but relatively high in Cantius. The low paraconid in the most primitive Teilhardina species suggests that the plesiomorphic euprimate condition was a low paraconid on $\mathrm{P}_{4}$.

## Postcrania

Talus. The talus of Teilhardina brandti is represented by two incomplete specimens (USNM 539577 and USNM 504329; Fig. 10A-E). USNM 539577 consists of a complete but damaged body and the lateral border of the head and neck, whereas USNM 504329 is a well-preserved astragalar body. As in other omomyids and T. belgica, the trochlear rims are parallel and sharp and the trochlea is shallowly grooved (Simpson, 1940; Godinot and Dagosto, 1983; Dagosto, 1988, 1993; Gebo, 1988; Covert and Hamrick, 1993; Anemone and Covert, 2000; Dunn et al., 2006). The medial malleolar surface is vertical. The lateral malleolar surface (fibular facet) is vertical superiorly and flares laterally plantarly, identical to the condition in T. belgica and similar to that in other omomyids. Specifically, the angle between the fibular
facet and the trochlea (= lateral tibial facet) measures 108 degrees (see measurement methods in Boyer et al., 2010). This is similar to values measured by Boyer et al. for Hemiacodon gracilis (107) and Tarsius syrichta (106). Anthropoids measured by Boyer et al. exhibit a more strongly vertical facet (96-102), while strepsirrhines and adapoids have more sloping facets (109-121). There is a large, deep fossa just dorsal to the ectal facet and posterior to the fibular facet, probably for attachment of the posterior talo-fibular ligament. The posterior shelf of the trochlea is moderately developed, similar to the condition in T. belgica (Szalay, 1976; Gebo, 1988). The ectal facet is tightly curved. The fragmentary talar neck and head of USNM 539577 indicate that the neck was long. The proximal aspect of the neck preserves a bony ridge delimiting the distal margin of a squatting facet for articulation with the distal tibia. This feature is common in omomyids (e.g., Simpson, 1940; Covert and Hamrick, 1993; Dunn, 2010).
Measurements (mm)—USNM 539577, body length: 2.40; estimated total length: 4.50; trochlear width: 1.90; total width: 2.50; ectal facet length: 1.60. USNM 504329, body length: 2.50; trochlear width: 1.90; total width: 2.55; ectal facet length: 1.60 .

Navicular. The navicular (USNM 542002, Fig. 10F-H) resembles those of cheirogaleids and omomyids in being longer than wide (Simpson, 1940; Gebo, 1988; Dagosto, 1993; Anemone and Covert, 2000; Dunn et al., 2006; Dunn, 2010). The navicular index (length/distal width $\times$ 100 ) is 165 -slightly below the range of extant cheirogaleids (166-295), slightly above the range for larger-bodied Malagasy strepsirrhines (66-162), and well above the range seen in modern platyrrhines (61-95). The navicular is significantly longer than that of Cantius (129) and Notharctus (110) but slightly shorter than that of other known omomyids (Hemiacodon $=176-190$; Ourayia $=$ 181-189; Chipetaia $=226$ ), and significantly less elongate than that of extant galagos (288-528) and tarsiers (434-626; ratios from Dunn, 2010). The proximal end is concave for articulation with the navicular facet of the talar head, with the medial side extending farther proximally than the lateral side. The distal end is relatively flat, lacking the expanded, bulbous articulations for the cuneiforms that are seen in galagos and tarsiers and more resembling the condition in other omomyids, adapids and strepsirrhines (Gebo, 1987; Dunn et al., 2006). The facet for the entocuneiform projects slightly more distally and is convex. A faint ridge is discernible separating the flat facet for the mesocuneiform from the slightly concave facet for the ectocuneiform.

Measurements (mm)—length: 3.80; distal width: 2.30.
Terminal phalanges. We attribute three isolated terminal phalanges (USNM 521825, 540587, and 542001; Fig. 11) to Teilhardina brandti based on their small size and distinctive euprimate morphology. The evolution of flat nails and expanded tactile pads (also called apical pads) on the tips of all digits is characteristic of euprimates and is widely considered a synapomorphy of the group (e.g., Clark, 1959; Cartmill, 1979, 1992; Dagosto, 1988, Hamrick, 1998; Soligo and Müller, 1999). This results in a unique morphology of the terminal phalanx in euprimates: the bone is flattened dorsoventrally and the tip is expanded mediolaterally. The expanded tip serves as the attachment for the flat nail dorsally and the tactile pad ventrally, and is often called the "apical tuft" (Hamrick, 1998, 2001; Mittra et al., 2007). The three


Fig. 10. Tarsal bones of Teilhardina brandti. A, USNM 539577, left talus in dorsal view (neck and head damaged). B-E, USNM 504329 , right talar body in dorsal (B), plantar (C), posterior (D), and lateral (E) views. F-H, USNM 542002, left navicular in dorsal (F), plantar (G), and distal (H) views. Note relatively elongate navicular and talar neck. ect, facet for ectocuneiform; ef, ectal facet; ent, facet for entocuneiform; ff, fibular facet; mes, facet for mesocuneiform; ps, posterior shelf; ptfl, posterior talofibular ligament; sf, squatting facet; tal, facet for talar head.
terminal phalanges reported here, which are the first known for Teilhardina, clearly exhibit an expanded apical tuft, indicating the presence of flat nails rather than claws. Differences in length and morphology-one is noticeably larger than the other two-probably indicate either that they are from different appendages (manus and pes), from different digits (hallux/pollex and lateral digits), or both. All have small nutrient foramina on the dorsal surface but no evident foramina on the volar surface.

The larger phalanx (USNM 542001) is more robust, being wider in all dimensions than the smaller ones. The tip of the phalanx is "arrow-shaped" in dorsal view, pointed at the apex with relatively straight sides rather than having a more generally rounded outline. One of the sides projects farther proximally than the other, making the apex appear asymmetrical. There are several nutrient foramina present on the apex of the phalanx. In distal view, the dorsal and ventral surfaces of the apical tuft are separated by a groove that continues from the apex of the phalanx proximally up the shaft and terminates before contact with the base. The shaft increases in width proximally; the base is significantly wider than the tip despite one side being broken. On the unbroken side there is a disto-laterally projecting flange of bone, probably for the insertion of the interosseus ligament. The extensor tubercle is well-developed. The sulcus for insertion of the deep flexor tendon on the ventral surface of the base is large and well-defined.

The smaller phalanges (USNM 521825, 540587) are more slender and more symmetrical than the larger one, having a more rounded apical region, a shaft of more uniform width, and a less expanded base. The extensor tubercle on the dorsal surface of the base is well-developed, but there is not a clearly defined flexor sulcus on the ventral surface. Distally, there is a faint ridge on the
ventral surface of the outer edge of the expanded tip of the phalanx. In lateral view both phalanges are dorsoventrally compressed, with a deeper base and tapering distally. This contrasts with the larger phalanx, which is more uniform in thickness.

The larger size, asymmetry, large flexor sulcus, and robust nature of the larger phalanx suggest that it may represent the terminal hallucal phalanx, as this is usually the largest and most robust ungual in extant primates and is often significantly larger in size than the others (Koenigswald, 1979; Godinot and Beard, 1991; Mittra et al., 2007). The smaller phalanges probably represent lateral digits either from the manus or pes. It has been suggested that terminal pedal phalanges have broader apical ends than those from the manus because feet have more morphological specializations for grasping (Godinot and Beard, 1991). However, a recent study indicated that there is no difference in shape between the terminal phalanges of the lateral digits in the manus and pes of extant prosimian primates (Mittra et al., 2007). In the Bridgerian adapoid Notharctus, the apical tufts of the terminal phalanges of manual digits III-V are slightly narrower than those of digits I and II (Hamrick and Alexander, 1996). The breadth of the apical tuft also varies by taxon, being more expanded in Cantius and Europolemur than in Smilodectes and Adapis (Covert, 1988; Godinot and Beard, 1991; Godinot, 1992). These observations further complicate the issue of allocating isolated phalanges to fore- or hind limbs. Compared to Bridgerian omomyid phalanges figured by Dagosto (1988), the small Wa-0 phalanges are narrower, with a less-expanded apical region, but this difference might be due to a more derived morphology in the Bridgerian taxa. The terminal phalanges of Teilhardina brandti differ from those of the adapoids for which lateral views of the unguals are known


Fig. 11. Distal phalanges of cf. Teilhardina brandti in lateral (A, D, G), palmar $(\mathbf{B}, \mathbf{E}, \mathbf{H})$, and dorsal $(\mathbf{C}, \mathbf{F}, \mathbf{I})$ views. AC, USNM 540587; D-F, USNM 521825; G-H, USNM 542001. at, apical tuft; et, extensor tubercle; fs, flexor sulcus; nf, nutrient foramina. Arrow indicates lateral groove.
(Europolemur, Adapis, Smilodectes, and possibly also Darwinius) in being flatter dorsoventrally (Koenigswald, 1979; Godinot and Beard, 1991; Godinot, 1992; Franzen et al., 2009). The groove on the tip of the larger phalanx has not been described in other fossil or extant primates, but is indicated in illustrations of adapid and omomyid phalanges (Koenigswald, 1979; Dagosto, 1988). Occurrence of the groove appears to be variable, and soft tissue correlates of this groove are unclear (Dagosto, personal communication).

Measurements (mm)—USNM 540587, $L=1.60$, proximal $W=1.10$, apical $W=0.70$; USNM $521825, L=$ 2.00, proximal $W=0.95$, apical $W=0.70$; USNM $542001, L=3.10$; apical $W=1.2$.

## RESULTS

## Principal coordinates analysis of $\mathbf{P}_{\mathbf{4}}$

Principal coordinates analysis of $\mathrm{P}_{4}$ variables resulted in $88 \%$ of the sample variance being captured by the first two coordinates (see Fig. 12). Coordinate 1 accounts for $78 \%$ of the sample variance. Adapoids and omomyids form separate clusters distinguished by their coordinate

1 values. Purgatorius, Palaechthon, and Altanius are outside of the morphospace defined by omomyids and adapoids, with Altanius essentially morphologically intermediate between the latter two clusters. Primitive members of the Teilhardina clade (T. asiatica and T. belgica) are closer morphologically to the plesiadapiforms, adapoids, and Altanius than are other Teilhardina species. While T. asiatica is outside of the T. belgica cluster, its morphology is not obviously "more basal" according to this phenetic analysis, as some specimens of T. belgica are closer to the outgroup taxa than is T. asiatica. The two measurable specimens of T. magnoliana occupy very different regions of the morphospace. The fragmentary CM 73229 is within the cluster defined by T. belgica, consistent with the hypothesis that T. magnoliana is a relatively basal member of the clade (Beard, 2008). However, CM 70434 plots as an outlier to the Teilhardina cluster, which we find telling given its qualitatively unusual appearance.

## Phylogenetic analysis

Two most parsimonious trees were recovered with a tree length of 169 steps, CI of 0.663 , and RI of 0.558 (calculated in PAUP). Tree topology is identical for these trees except for the placement of Teilhardina asiatica and T. magnoliana. In the first most parsimonious tree, Teilhardina asiatica is the most basal species of Teilhardina (as in previous phylogenetic analyses: Ni et al., 2004; Smith et al., 2006; Beard, 2008) and is the sister taxon to a clade consisting of all other omomyids in the analysis; T. magnoliana + T. belgica are sister taxa and together form a clade that is sister to the other omomyids (Fig. 13A). T. asiatica and T. magnoliana switch positions in the other most parsimonious tree (Fig. 13B). In both trees T. brandti is more derived than T. asiatica, T. belgica, and T. magnoliana, and less derived than T. americana. A strict consensus (Fig. 13C) results in a polytomy at node 5 , consisting of T. asiatica, T. belgica, T. magnoliana, and a clade consisting of remaining omomyids, in which T. brandti is most primitive. Character support for each node is listed in Figure 13C.

Our results, like other recent analyses of Teilhardina (Smith et al., 2006; Beard, 2008; Tornow, 2008), suggest that Teilhardina is paraphyletic (see also Rose and Bown, 1993) or possibly even polyphyletic. This is perhaps not surprising considering its basal position among omomyids. These possibilities should be further explored in future phylogenetic analyses of primitive euprimates.

## DISCUSSION

Teilhardina brandti is the oldest and most primitive omomyid for which incisor alveoli are adequately preserved to enable a judgment of their relative size and orientation. The incisors were unequivocally small, with essentially vertical roots. Although $I_{1}$ was slightly larger than $I_{2}$, it was neither procumbent nor enlarged like $I_{1}$ in North American Teilhardina crassidens, T. americana-T. crassidens intermediates, T. demissa, Anemorhysis, Tetonius, Pseudotetonius, and Steinius. Early Eocene Altanius orlovi shows similar compression of antemolar teeth, but it has a two-rooted $P_{2}$ and apparently had small anteriorly inclined incisors whose alveoli are about equal in size. The incisors of Donrussellia are unknown, but the adapoid Cantius has uncompressed antemolar teeth and


Fig. 12. Plot of principal coordinates 1 and 2 resulting from principal coordinates analysis of four variables of $P_{4}$ morphology (buccolingual angle, mesial angle, preprotocristid verticality angle, trigonid width index). See Methods and Results for further explanation and Appendix D for values used. Numbered groups are: 1, omomyids; 2, notharctids; 3, asiadapines; 4, Teilhardina belgica; 5, Teilhardina brandti; 6, Teilhardina americana; 7, Teilhardina crassidens; 8, Tetonius matthewi. Note that Teilhardina asiatica, T. belgica, and one specimen of T. magnoliana plot closer to the region including plesiadapiforms and notharctids, consistent with cladistic results and previously described morphoclines indicating that these forms are more basal.
small, strongly inclined incisors, with $\mathrm{I}_{2}$ larger than $\mathrm{I}_{1}$. These observations provide evidence that the plesiomorphic condition in euprimates was to have small incisors, but their orientation is still uncertain. Hypertrophy of $I_{1}$ in various omomyids was, therefore, a derived condition.

The Teilhardina brandti sample described here, in comparison with other primitive euprimates, provides further evidence that the following character states are plesiomorphic for Omomyidae and probably euprimates as well: four premolars including $\mathrm{P}_{1}$ that is not laterally displaced; $\mathrm{P}_{4}$ with small and low paraconid and metaconid; simple upper molars in which the hypocone and nannopithex fold are absent and the lingual cingulum is incomplete, and smooth enamel. Uncompressed antemolar dentition including an unreduced canine and a tworooted $P_{2}$ are probably also primitive for euprimates.

Comparison of the postcanine teeth among species of Teilhardina indicates that T. brandti is morphologically intermediate between T. belgica and T. americana, and belongs in the genus Teilhardina. While its molar size more closely approximates that of T. americana, many features of the lower cheek teeth, including metaconid height of $\mathrm{P}_{4}$ and proportions of $\mathrm{M}_{1}$, are more similar to T. belgica than to T. americana or are intermediate between the two species (Figs. 6 and 14, Table 3). The upper teeth are also more similar morphologically to those of T. belgica than to those of T. americana, although one specimen (UF 244460) shows features more typical of T. americana (crenulation, a lingual cingulum,
and a weak nannopithex fold), suggesting a transitional stage. An alternative interpretation of this specimen is that it indicates the presence of T. americana in addition to T. brandti during the PETM; however, it is very improbable that two so similar species coexisted at that time, when all other evidence suggests they are segments of the same anagenetic lineage.
The isolated foot bones we attribute to Teilhardina brandti are among the oldest known for Euprimates. The terminal phalanges, which are also the smallest known for Euprimates, attest to the presence of nails rather than claws and support the hypothesis that nails were present in the last common ancestor of living euprimates (Soligo and Müller, 1999). However, the diminutive size of T. brandti and other basal euprimates is inconsistent with Soligo and Müller's contention that nails arose in association with an increase in body size early in the evolution of Euprimates (see also Gebo, 2004, who argued that the ancestral euprimate must have been very small, and Bloch et al., 2007, and Sargis et al., 2007, who argued that nails evolved in the common ancestor of plesiadapoids and Euprimates). The relative elongation of the navicular compared to that of notharctids, and the long, straight talar neck, are evidence that very early in their history omomyids were already more specialized for active arboreal locomotion than were notharctids.

The cladistic analysis presented here is the most comprehensive attempt to resolve interrelationships among


Fig. 13. Results from cladistic analysis of 47 characters and 13 taxa (see Appendix). A, One of two most parsimonious trees resulting from branch and bound search using unordered characters; tree length: 169. B, Second of two most parsimonious trees. C, Strict consensus of the two most parsimonious trees. Unambiguous synapomorphies, with superscript R indicating instances of reversal: Node $1,3(1), 8(1), 15(1), 17(1), 33(1)$, 38(3), 42(1); Node 2, 4(1), 5(1), 16(1), 45(1); Node 3, 10(1), 28(2), 37(2), 38(2); Node 4, 12(2), 30(2), 43(1); Node 5, 31(1), 42(0) ${ }_{\mathrm{R}}$; Node 6, 11(1), 28(1); Node 7, 3(0) R, 18(0) ${ }^{\text {R }}, 29(2), 37(1), 45(0){ }^{R}$; Node 8, 1(1), 20(1), 21(1); Node 9, 22(2), 26(2), 29(3), 46(2).
species of Teilhardina to date. It incorporates upper tooth characters for the first time, as well as additional characters of the lower dentition, some of which quantify previous qualitative observations. Postcranial traits were excluded because they are still much too poorly known to be informative within Teilhardina. Strict consensus of the two most parsimonious trees results in a polytomy among the most basal species of Teilhardina, whereas the two most parsimonious trees recover either T. asiatica or T. magnoliana as the most basally divergent species of Teilhardina. Previous cladistic analyses have also resulted in T. asiatica as the most basally divergent species of Teilhardina (e.g., Ni et al., 2004; Smith et al., 2006; Beard, 2008). Morphological features of T. asiatica, such as the less-reduced $P_{1}$ relative to other species of Teilhardina, suggest that this is the most primitive species known (e.g., Ni et al., 2004). Although features of the anterior dentition cannot be assessed for T. magnoliana, the only other cladistic analysis that incorporated T. magnoliana and T. asiatica supported T. asiatica as more basally divergent (Beard, 2008). Consequently, we regard the topology of the first most parsimonious tree (Fig. 13A), with T. asiatica basal, as the most likely arrangement based on present evidence.

Carbon isotope stratigraphy suggests that the relative ages of T. asiatica, T. belgica, and T. brandti are sequential and that all were restricted to the PETM. Based on this evidence and stage of evolution, Smith et al. (2006) hypothesized that Teilhardina originated in Asia and dispersed westward, successively to Europe and then to North America. Beard (2008), however, used sequence stratigraphy to argue that Teilhardina magnoliana, from the uppermost Tuscahoma Formation Red Hot local fauna of Mississippi, predated both T. brandti from the Bighorn Basin and T. belgica from the Tienen Formation at Dormaal, Belgium. On the basis of this inference, he postulated that dispersal was in the opposite directionfrom Asia to North America (Gulf Coast) at the beginning of the PETM, and from there northward to Wyoming and eastward to Europe. He bolstered his proposal by citing various mammalian species in the Red Hot fauna that resemble species from the very short Wa-M biozone that immediately underlies Wa-0 in the Bighorn Basin. However, most of the taxa in the Red Hot fauna that are comparable to Bighorn Basin taxa, including the rodent Tuscahomys (one of the two taxa Beard mentioned), are similar to species known from throughout the Wa-0 interval. One Tuscahoma species considered particularly suggestive of a close correspondence with the Wa-M biozone is the tiny condylarth Haplomylus meridionalis (Beard and Dawson, 2009), which was compared to a species (H. zalmouti) known only from Wa-M in the Bighorn Basin. However, Gingerich (2010) showed that $H$. meridionalis is much closer in size to early Wasatchian H. speirianus, suggesting on this basis that H. meridionalis is more comparable to Wa-1 species.

The Red Hot local fauna comes from a vertebrate lag at the top of the Tuscahoma Formation immediately below the Wasatchian Bashi Formation, and recent studies of pollen and foraminifera suggest that the vertebrate lag dates from later rather than earlier in the PETM (Harrington, 2003; Fluegeman, 2009). The conclusion that the Red Hot local fauna dates from well after the beginning of the PETM also appears to be supported by sedimentary evidence from a recent core taken near the site, which found the Paleocene-Eocene boundary to be 30 feet below the vertebrate lag (Dockery and


Fig. 14. Proportions and size (ln area) of $\mathrm{M}_{1}$ in Teilhardina in the lower Willwood Formation of the Bighorn Basin, Wyoming. Stratigraphic levels (m) of T. brandti from the Sand Creek Divide section (SCD) are indicated at the lower left of each plot; three specimens from the Cabin Fork section have been interpolated based on Figure 1. Levels in other sections in the southern Bighorn Basin (BHB; see Bown et al., 1994) are shown at right of each plot. The 30-m level in the latter sections appears to correlate closely with $\sim 45 \mathrm{~m}$ in the SCD section. T. belgica and T. asiatica are plotted stratigraphically below T. brandti, following Smith et al. (2006). T. magnoliana is shown below the section, as its stratigraphic position is ambiguous. Note that $\mathrm{M}_{1}$ increases in relative width through the section, but $\mathrm{M}_{1}$ area shows no obvious trend, though T. asiatica, T. belgica, and T. magnoliana average smaller than the Bighorn Basin species. Data from Bown and Rose (1987), Smith et al. (2006), Beard and Dawson (2009), and this paper. Data for T. belgica based on original measurements of epoxy casts of specimens listed in Appendix C.

Thompson, 2009). In the absence of detailed chemostratigraphy of the Tuscahoma Formation, the age of T. magnoliana relative to other early species of Teilhardina is difficult to establish; but existing evidence casts doubt on the hypothesis that the Red Hot fauna and T. magnoliana predate the first occurrence of Teilhardina in the Bighorn Basin. It may not be possible to determine the relative age of the Red Hot local fauna compared to the Bighorn Basin Wa-0 faunas without knowing its precise position relative to the CIE, and specifically, to the minimum value of $\delta^{13} \mathrm{C}$. A detailed carbon isotope section through the Tuscahoma Formation, which could resolve the controversy, has not yet been published.
T. magnoliana appears to be as primitive as T. brandti or more so in most features, but possibly more derived in others (e.g., $\mathrm{P}_{4}$ metaconid height and talonid structure). Judging from dental morphology, it is more likely that $T$. magnoliana evolved independently from T. belgica than directly from either T. asiatica or T. brandti, but a larger sample and more complete specimens are needed in order to determine its phylogenetic position with confidence. Consequently, available evidence favors the westward dispersal hypothesis of Smith et al. (see also Gingerich et al., 2008).

## CONCLUSIONS

1. The first known upper teeth of Teilhardina brandti have primitive molars with no hypocone or nannopithex fold, an incomplete lingual cingulum, and smooth enamel. They are essentially identical to upper teeth of T. belgica but slightly larger.
2. A new dentary reveals the lower dental formula (2.1.4.3), including alveoli of small, apparently vertical incisors ( $\mathrm{I}_{1}$ slightly larger than $\mathrm{I}_{2}$ ), a moderatesized canine, a reduced and laterally displaced $\mathrm{P}_{1}$, and a single-rooted $P_{2}$. Other new specimens show talonid variation in $\mathrm{P}_{4}$.
3. Analysis of $P_{4}$ characters confirms the low metaconid on $\mathrm{P}_{4}$ of Teilhardina brandti (similar in height to that of T. belgica). Molar lengths are comparable to those of T. americana, whereas relative molar width is comparable to that of T. belgica.
4. The first known postcrania for Teilhardina brandti include tarsals (elongate talus and navicular, indicative of active arboreal locomotion) and terminal phalanges (the oldest evidence for nails in euprimates).
5. Phylogenetic analysis places Teilhardina brandti essentially intermediate between T. belgica and
T. americana. Two most parsimonious trees were recovered. As in previous analyses, the first result supports T. asiatica as the most basal species of Teilhardina. The second result supports T. magnoliana as the most basal species, but we consider this scenario less likely in light of the available geological and morphological evidence.
6. T. magnoliana is probably not older (and quite possibly is younger) than T. brandti from the Bighorn Ba$\sin$. Together with the morphologic evidence, this suggests that the hypothesis that Teilhardina dispersed from Asia westward to Europe and then to North America during the PETM remains most probable.

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|  | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | 13 | 14 | 15 | 16 | 17 | 18 | 19 | 20 | 21 | 22 | 23 | 24 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| D. provincialis | 0 | 1 | 1 | 1 | 1 | 1 | 2 | 1 | 0,1 | 1 | 0 | 2 | 0 | 0 | 1 | 1 | 1 | 0,1 | ? | 0 | 0 | 0 | 0 | 0 |
| D. gallica | 0 | 1 | 1 | 1 | 1 | 0 | 0 | 0 | 0 | ? | 1 | 2 | 1 | 0 | 0 | 1 | 1 | 0,1 | ? | 0 | ? | ? | ? | ? |
| T. asiatica | 0 | ? | 1 | 1 | 1 | 1 | ? | 1 | 0 | $?$ | ? | ? | ? | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 0 | 0 | 1 | 0 |
| T. belgica | ? | 0 | 1 | 1 | 1 | 1 | 0 | 1 | 0 | $?$ | 0 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 0 | 1 | 1 | 1 |
| T. brandti | 0 | 0 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | ? | 1 | 0 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 0 | 1 | 1 | ? |
| T. magnoliana | 0 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 0 | ? | 0 | 0 | 0,1 | 1 | 1 | 1 | 1 | 1 | ? | 0 | ? | ? | ? | ? |
| T. americana | 1 | 0 | 0 | 1 | 1 | 1,2 | 1,2 | 0,1 | 1 | 1 | 1 | 0 | 1 | 1 | 0,1 | 1 | 0 | 0 | 1 | 1 | 1 | 1 | 1 | 1 |
| T. crassidens | 1 | 0 | 0 | 1 | 1 | 2 | 2 | 1 | 1,2 | 1 | 1 | 0 | 0,1 | 1 | 1 | 1 | 0 | 0 | 1 | 1 | 1 | 2 | 1 | 1 |
| T. matthewi | 1 | 0 | 1 | 1 | 1 | 1,2 | 0 | 1 | 1 | 1 | 1 | 0 | 1 | 1 | 0,1 | 1 | 0 | 0 | 1 | 1 | 1 | 2 | 1 | 0 |
| S. vespertinus | 0 | 0 | 0 | 1 | 1 | 1 | 0,1 | 1 | 2 | 1 | 1 | 1 | 1 | 0 | 1 | 1 | 0,1 | 0 | 1 | 0 | 0 | 1 | 1 | 1 |
| Purgatorius | 0 | 0 | 0 | 0 | 0 | 1 | 0 | 0 | 0,1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | ? | 0 | 0 | 0 | 0 | 0 |
| Palaechthon | 0 | 1 | 1 | 0 | 0 | 1 | 0 | 1 | 2 | 0 | 0 | 1 | 0 | 0 | 1 | 0 | 1 | 0 | 0 | 0 | 0,1 | 2 | 1 | 1 |
| Altanius | 1 | 0 | 1 | 1 | 1 | 0 | 0 | 1 | 2 | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 1 |
|  | 25 |  | 26 | 27 | 28 | 29 | 30 | 31 | 32 | 33 | 34 | 35 | 36 | 37 | 38 | 39 | 40 | 41 | 42 | 43 | 44 | 45 | 46 | 47 |
| D. provincialis | 1 |  | ? | 1 | ? | ? | 2 | ? | 1 | 1 | 0 | 1 | 1 | 2 | 2 | 2 | 0 | 1 | 1 | 1 | ? | ? | ? | ? |
| D. gallica | ? |  | 0 | 1 | 0 | 0 | 2 | 0 | 1 | 1 | 1 | 0 | 0 | 1 | 2 | 2 | 0 | 0 | 0,1 | 1 | ? | ? | 1 | 2 |
| T. asiatica | 0 |  | 0 | 2 | 1 | 0 | 0 | 1 | 0 | 1 | 0 | 0,1 | 0 | 2 | 0 | 1 | 1 | 1 | 0 | 0 | ? | ? | 0 | 2 |
| T. belgica | 0 |  | 1 | 1,2 | 1 | 1 | 1 | 1 | 0 | 0 | 0 | 1 | 1 | 2 | 1 | 2 | 1 | 1 | 0 | 0 | 1 | 1 | 0 | 2 |
| T. brandti | ? |  | 1 | 2 | 1,2 | 1 | 1 | 1 | 0 | 0,1 | 0,1 | 1 | 1 | 2 | 1 | 2 | 0,1 | 1,2 | 0 | 0 | 1 | 1 | 0 | 1 |
| T. magnoliana | ? |  | 1 | 2 | 2 | 1 | 1 | 1 | 0 | 0 | 0 | 1 | 0 | 2 | 2 | 2 | 0 | 0 | 0 | 0 | ? | ? | 0 | ? |
| T. americana | 1 |  | 1 | 2 | 2 | 2 | 0,1 | 1 | 0 | 1 | 1 | 2 | 2 | 1 | 2 | 2 | 1 | 1 | 0 | 0 | ? | ? | 0 | 1,2 |
| T. crassidens | 1 |  | 2 | 1 | 3 | 3 | 0,1 | 1 | 0 | 1 | 1 | 3 | 3 | 0 | 2 | 2 | 2 | 2 | 0 | 0 | 1 | 0 | 2 | 0 |
| T. matthewi | 1 |  | 2 | 2 | 2 | 3 | 1,2 | 1 | 0 | 0,1 | 1 | 3 | 3 | 1 | 3 | ? | 1,2 | 2 | 0 | 0 | 1 | 0 | 2 | 1 |
| S. vespertinus | 1 |  | 1 | 2 | 2 | 2 | 1 | 1 | 0 | 1 | 1 | 1 | 1 | 1 | 2,3 | ? | 1 | 2 | 0,1 | 1 | 1 | 0 | 0 | 0 |
| Purgatorius | 0,1 |  | 0 | 0 | ? | 0 | 1 | 0 | 0,1 | 0 | 0 | 2 | 0,2 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | ? |
| Palaechthon | 1 |  | 0 | 0 | 2 | 2 | 0,1 | 0 | 0 | 1 | 0 | 1 | 1 | 0 | 3 | 0 | 0 | 0 | 1 | 1 | 0,1 | 0 | 0 | 0 |
| Altanius | 1 |  | 1 | 0 | 3 | 0 | 1 | 0 | 1 | 1 | 0 | 2,3 | 1,2,3 | 0 | 3 | 1 | 0,1 | 2 | 1 | 0 | 1 | 1 | 1 | 0 |

${ }^{\text {a See Supplemental information for Nexus file. }}$

## APPENDIX B: CHARACTER DESCRIPTIONS

Description of dental characters used in the cladistic analysis, with comments on characters. ${ }^{1}$ Upper molar characters mainly adapted from Seiffert et al. (2005). Lower tooth characters mainly adapted from Smith et al. (2006; indicated by S) or Beard (2008; indicated by B). N designates new characters, while (m) indicates that character descriptions or states have been modified to facilitate scoring and repeatability.

1. $P^{4}$ parastyle: (0) present; (1) very small to absent 2. $\mathrm{P}^{4}$ metastyle: (0) absent; (1) present
2. $\mathrm{P}^{3-4}$ buccal cingulum: (0) strong; (1) absent or weak

[^1]4. $\mathrm{P}^{4}$ metacone: (0) present; (1) absent
5. $\mathrm{P}^{4}$ protocone: (0) high relative to paracone; (1) low relative to paracone
6. $\mathrm{P}^{4}$ occlusal outline: (0) protocone lobe mesiodistally compressed, about half L of buccal margin (buccal $\mathrm{L} /$ lingual $\mathrm{L}<0.60$ ); (1) protocone lobe about $2 / 3 \mathrm{~L}$ of buccal margin (0.61-0.70); (2) >3/4 L of buccal margin ( $>0.71$ )
7. $\mathrm{N} . \mathrm{M}^{2}$ parahypocrista: (0) absent; (1) incipient; (2) distinct
8. $\mathrm{M}^{1}$ hypocone size: (0) present; (1) absent
9. $\mathrm{M}^{1-2}$ nannopithex fold: (0) absent; (1) weak; (2) strong
10. N. Position of nannopithex fold: (0) posterolingually directed, to postcingulum; (1) posteriorly or posterobuccally directed, does not join cingulum
11. $\mathrm{M}^{1-2}$ premetaconule crista: (0) absent or weak; (1) strong
12. $\mathrm{M}^{1-2}$ postmetaconule crista: (0) strong; (1) weak; (2) absent
13. $\mathrm{M}^{1-2}$ lateral posterior transverse crista: (0) indistinct; (1) distinct
14. $\mathrm{M}^{2}$ shape (W/L): (0) wide (1.3-1.65); (1) very wide ( $>1.65$ )
15. $\mathrm{M}^{2}$ hypocone: (0) present; (1) absent
16. $\mathrm{M}^{2}$ postmetacrista: (0) longer, trenchant, and more buccally extended; (1) short, indistinct or absent
$17 . \mathrm{M}^{2}$ buccal extension of paracone: (0) slight expansion; (1) no expansion
18. $\mathrm{M}^{1-3}$ anterior cingulum: (0) strong, complete, long; (1) strong, short
19. $\mathrm{M}^{3}$ paraconule: (0) distinct, somewhat large (1) absent or very small
20. N. Enamel texture: (0) smooth; (1) crenulated
21. $\mathrm{S}(\mathrm{m}$ ). Lower canine alveolus: ( 0 ) distinctly larger than adjacent teeth; (1) not distinctly larger than adjacent teeth
22. $\mathrm{S} . \mathrm{P}_{1}$ : (0) present; (1) reduced and laterally shifted; (2) absent
23. $\mathrm{B}(\mathrm{m}) . \mathrm{P}_{2}$ : (0) two-rooted, unreduced; (1) one-rooted and reduced, or absent
24. $\mathrm{S}(\mathrm{m})$. $\mathrm{P}_{3-4} \mathrm{~L}: \mathrm{M}_{1-2} \mathrm{~L}$ : (0) $\mathrm{P}_{3-4}$ not compressed $\left(\mathrm{P}_{3-4}\right.$ $>0.85 \times \mathrm{M}_{1-2} \mathrm{~L}$ ); (1) $\mathrm{P}_{3-4}$ anteroposteriorly com-$\operatorname{pressed}\left(\mathrm{P}_{3-4}<0.85 \times \mathrm{M}_{1-2} \mathrm{~L}\right)$

Compression of premolars is often said to characterize various lineages of omomyids, in contrast to adapoids. Although this character was used previously by Smith et al. (2006) and Beard (2008), it was not quantified. Here length of the teeth involved was compared (means were scored where sufficient specimens were available), with in some cases surprising results. Using this criterion, scores in many cases differ from previous assessments. Apparent compression of $\mathrm{P}_{3-4}$ may be due in part to relative widening of these teeth. In addition, the relative length is affected by mesiodistal compression of the molars, which may explain why Tetonius scores as having less compressed premolars than Steinius. Antemolar compression is more evident anterior to $\mathrm{P}_{3}$.
25. B. $\mathrm{P}_{3}$ protoconid: (0) mesial, preprotocristid relatively vertical (mesial to the midpoint of the trigonid); (1) approximately at midpoint or distal to midpoint, with more gently sloping preprotocristid.

This character (character 4 of Beard, 2008) replaces character 3 of (Smith et al. 2006: $\mathrm{P}_{3}$ shape, described as relative procumbency), allowing it to be assessed more objectively. The trigonid was viewed lingually and protoconid position was observed along a line from the mesial end of the trigonid to the talonid notch.
26. N. $\mathrm{P}_{4}$ buccolingual angle: $(0)<39$; (1) 39-55; (2) $>55$

The tooth is oriented by using the lingual margin to define the mesiodistal axis. The occlusal surface is oriented so the lingual and buccal faces of the protoconid have equal slopes and so the postvallid surface and preprotocristid surface have roughly equal slopes. The tooth is then viewed from the mesial end. The buccal arm of the angle is defined by the buccal surface of the protoconid. The lingual arm is defined by the lingual surface, excluding any projection of the metaconid.
27. $\mathrm{S}(\mathrm{m}) . \mathrm{P}_{4}$ paraconid: (0) distinct and high; (1) distinct and low; (2) small and low, very small, or absent
28. N. $\mathrm{P}_{4}$ metaconid height 1: metaconid height/protoconid height: (0) <0.60; (1) 0.60-0.68; (2) 0.69-0.78; (3) >0.78
$\mathrm{P}_{4}$ metaconid height has long been recognized as a critical morphoclinal character in Teilhardina (Bown and Rose, 1987; Smith et al., 2006; Beard, 2008), yet it has previously been assessed only qualitatively. In an effort to characterize metaconid height more objectively, we have quantified it in two ways. For character 28 , the tooth was viewed lingually and a base line was drawn joining the lowest extent of the enamel of the trigonid and talonid. Height of the metaconid was measured as a perpendicular from this base line to the tip of the metaconid. Character 28 is the index of this value compared to protoconid height ( $=$ crown height) measured from the same base line. See also character 47.

Specimens with obvious wear or damage to the protoconid were excluded.
29. N. $\mathrm{P}_{4}$ preprotocristid verticality angle (0) $<43$; (1) 43-52; (2) 53-62; (3) >62

The tooth is oriented using the same procedure as for character 26 . The tooth is then viewed from the buccal side. The mesial arm of the angle is defined by the greatest linear segment of the preprotocristid, which is usually the most mesial part that contacts the paraconid, when present. The distal arm of the angle is defined by the postvallid surface of the trigonid, excluding any topography due to the projection of the metaconid.
30. S. $\mathrm{P}_{4}$ cristid obliqua: (0) extends to base of trigonid wall; (1) runs part way up trigonid wall; (2) extends up trigonid wall above metaconid
31. N. $\mathrm{P}_{4}$ trigonid width index ( 0 ) <1.21; (1) 1.21 and greater

The index used is trigonid length/trigonid width. Trigonid length was used instead of total length so that CM 73229 (Teil. magnoliana) could be included.
32. $\mathrm{S} . \mathrm{M}_{1}$ cristid obliqua: (0) extends to base of trigonid wall or runs part way up trigonid wall; (1) extends to metaconid
33. S. Buccal cingulids on lower molars: (0) weak or absent; (1) moderate to strong

Buccal cingulids in some taxa (e.g., Teilhardina asiatica) are strong on parts of some molars but weak or absent elsewhere. Such taxa were scored as moderate.
34. S . Hypoconulids on $\mathrm{M}_{1-2}$ : (0) well defined; (1) weak

Hypoconulids were scored as well defined if either a distinct cusp is present or a distinct elevation occurs on the postcristid in the position of a cusp.
35. $\mathrm{B}(\mathrm{m}) . \mathrm{M}_{1}$ shape: (0) narrow ( $\mathrm{M}_{1} \mathrm{~L} / \mathrm{W}>1.40$ ); (1) somewhat square ( $\mathrm{M}_{1} \mathrm{~L} / \mathrm{W}=1.30-1.40$ ); (2) more nearly square ( $\mathrm{M}_{1} \mathrm{~L} / \mathrm{W}=1.20-1.30$ ); (3) square and inflated ( $\mathrm{M}_{1} \mathrm{~L} / \mathrm{W}<1.20$ )
$\mathrm{M}_{1}$ shape is included here, following Beard (2008), although in most cases it duplicates character 36 ( $\mathbf{M}_{2}$ shape).
36. $\mathrm{S}(\mathrm{m}) . \mathrm{M}_{2}$ shape: (0) narrow $\left(\mathrm{M}_{2} \mathrm{~L} / \mathrm{W}>1.30\right)$; (1) somewhat square ( $\mathrm{M}_{2} \mathrm{~L} / \mathrm{W}=1.21-1.30$ ); (2) more nearly square $\left(\mathrm{M}_{2} \mathrm{~L} / \mathrm{W}=1.11-1.20\right)$; (3) square and inflated $\left(\mathrm{M}_{2} \mathrm{~L} / \mathrm{W}<1.10\right)$

The single known complete $\mathrm{M}_{2}$ of T. magnoliana (the holotype) is narrow ( $L / W=1.32$ ), whereas $\mathrm{M}_{2} \mathrm{~S}$ of T. belgica and T. brandti are, on average, wider $(L / W$ index $=1.25$ for both species). However, three specimens of T. brandti have indices of 1.30-1.34, comparable to T. magnoliana.
37. $\mathrm{S}(\mathrm{m}) . \mathrm{M}_{2}$ entoconid: (0) High (as high as or nearly as high as hypoconid); (1) intermediate; (2) Low (distinctly lower than hypoconid in lingual and posterior views)

As for many characters, while extremes are simple to code, many specimens fall into a more or less intermediate state that is difficult to code consistently.
38. $\mathrm{S}\left(\mathrm{m}\right.$ ). $\mathrm{M}_{3}$ hypoconulid lobe: ( 0 ) very narrow ( $<1 / 3$ as wide as $\mathrm{M}_{3}$ maximum width); (1) narrow ( $36-45 \%$ as wide as $\mathrm{M}_{3}$ maximum width); (2) intermediate (46$55 \%$ as wide); (3) wide ( $>56 \%$ as wide)

Variations in shape of the $\mathrm{M}_{3}$ hypoconulid lobe make it difficult to obtain consistent width measurements. As a measure of hypoconulid lobe size, maximum width of the hypoconulid lobe was compared with maximum $\mathrm{M}_{3}$ width; higher values reflect relatively wider hypoconulid lobes. Values for Teilhardina species are: T. asiatica $=0.32$; T. brandti $=$ 0.39 ; T. belgica $=0.41$; T. magnoliana $=0.47$; T. americana $=0.51 ;$ T. crassidens $=0.54$.
39. N. $\mathrm{M}_{2}$ relief index: ( 0 ) $>0.55$; (1) $0.55-0.53$; (2) $<0.53$ The relief index was calculated as the [natural logarithm (square root of $\mathrm{M}_{2} 3$-D surface area/square root of $\mathrm{M}_{2}$ 2-D surface area)]. See Boyer (2008) for measurement methods and descriptions.
40. $B(m) . M_{2}$ paraconid position: (0) well separated from metaconid; (1) intermediate; (2) connate with metaconid or absent.
41. $B(m) . M_{2}$ paraconid height: (0) situated low on trigonid, well below metaconid; (1) slightly below metaconid; (2) situated high on trigonid, at or near level of metaconid.
42. $\mathrm{B}(\mathrm{m}) . \mathrm{M}_{3}$ trigonid: (0) wider than talonid; (1) similar in width to talonid
$\mathrm{M}_{3}$ trigonid was scored as similar in width to the talonid if differences in width were $\leq 0.05 \mathrm{~mm}$; if the difference was $\geq 0.10 \mathrm{~mm}$ the trigonid was scored as wider than the talonid. (Measurements were taken to the nearest 0.05 mm ).
43. $\mathrm{B}(\mathrm{m}) . \mathrm{M}_{3}$ trigonid: (0) significant baso-buccal distension; (1) buccal surface nearly vertical

This character ( $=$ character 24 of Beard, 2008) is considered to include Beard's character 25, relative inflation of $\mathrm{M}_{3}$ trigonid cusps.
44. N. Lower incisors (number): (0) 3; (1) 2

Silcox (2001) inferred the presence of $\mathrm{I}_{3}$, and thus three lower incisors, in Palaechthon nacimienti, but only two in P. alticuspis.
45. N. Size of $\mathrm{I}_{1}$ : (0) $>\mathrm{C}$ (hypertrophied); (1) $<\mathrm{C}$ (comparable to other incisors)
46. N. $\mathrm{P}_{4}$ mesial angle: (0) $125-145$; (1) $>145$; (2) $<125$

The tooth is oriented as for character 26 and is viewed buccally. The upper (more dorsal) arm of the angle is the same as the mesial arm of the verticality angle defined for character 29. The lower (more ventral) arm is defined by the longest linear segment of the anterior part of the trigonid's cervical margin. This segment usually extends from the mesial half of the mesial root to the end of the anteroexternal cingulid.
47. N. $P_{4}$ metaconid height 2 : metaconid height $/ P_{4}$ length: (0) $>0.66$; (1) $0.56-0.66$; (2) $<0.56$

Character 47 compares metaconid height to $\mathrm{P}_{4}$ mesiodistal length. The tooth is oriented as for character 26 and is viewed lingually. Metaconid height is the dorsoventral distance (perpendicular to the occlusal plane) from the apex of the gap between the mesial and distal roots to the notch between the protoconid and metaconid.

## APPENDIX C: TAXA AND SPECIMENS EXAMINED, WITH COMMENTS

Teilhardina asiatica-Cast of IVPP V-12357 (lower); other data from Ni et al. (2004).

Teilhardina belgica-Casts of IRSNB nos. M64 (lectotype), M65, M4290, M4291, M4292, M4296, CL 182, CL231, CL246, CL455, CL457, WL13, WL128, WL159, WL180, WL1052, WL1060, WL1062, WL1180, WL1299, WL1398; also casts of RS nos. DIII15S, DIII16S, DIII19S, DIII20S, DIII467, DIII844, DIII952, DIIC761. Other data from Smith et al. (2006).

Teilhardina brandti-UM 99031 (holotype, cast), USNM nos. 493913, 493914, 521795 525543-525546, 525621, 525622, 533494, 533505, 533554, 538082, 538084, 538361, 539466, 539467, 540598; UF nos. 244451, 244453-244458, 244460.

Teilhardina magnoliana-Casts of CM nos. 67854, 67856, 67858, 67860, 67861, 70422, 70427, 70430, 70431, 70433, 70434, 70435 (holotype), 70436, 73229, 77209-77212.
$\mathrm{P}_{4}$ morphology, particularly changes in width and metaconid development, are of particular significance in the evolution of this genus (e.g., Rose and Bown, 1984; Bown and Rose, 1987). Unfortunately, only three specimens of $\mathrm{P}_{4}$ are known for T. magnoliana, one badly eroded and the other two fragmentary trigonids. Consequently $\mathrm{P}_{4}$ traits are difficult to assess with certainty and appear to differ significantly between the two specimens we examined (CM 70434 and 73229). This ambiguity may affect the outcome of the analysis.
Teilhardina americana-UF nos. 244452, 244459; UM nos. $65770,75610,76600$ (casts); USGS nos. 3849, 3863, 8886, 9037, 12747, 15406, 15450, 23838, 23963, 25324 (cast); USNM nos. 539489, 539491; UW nos. 6896 (holotype, cast), 6907, 7098 (cast), 8871, 8961. Other data from Bown and Rose (1987).
Teilhardina crassidens-UM 71071 and 73908 (casts); USGS 7204, 15409; USNM 487869; UW 8959 (holotype, cast); YPM 24626 and 30721 (casts). Other data from Bown and Rose (1987).

Tetonius matthewi-USNM nos. 487864, 487865, 487870, 521477, 527712, 527713, 533455, 533620; USGS 3841; CM 12190 (holotype, cast); YPM 23031 and 25027 (casts); UM 76675 (cast). AMNH 4194, holotype of $T$. homunculus, was also compared and was found to score the same in nearly all features.
This species was used because it is much better known than the slightly more primitive T. mckennai.
Steinius vespertinus-USGS 502 (cast); USNM nos. 491935-491954, 527694.
Altanius orlovi-Casts of PSS nos. 7/20-8 (holotype), 20-58, 20-61, 20-85.
Donrussellia provincialis-Casts of MNHN RI-170 (holotype), RI-229. Other data from Godinot (1981, 1998).
Donrussellia gallica-Casts of MNHN Av nos. 4562, 4598, 4603, 4613, 4618, 4711, 4714, 4774, 4830, 48544856, 5015, 5017, 5022, 5654, 5721, 5755 (holotype), 5765, 5767, 5795, 5807, 5841, 5846, 5859, 5873, 6757, 6848, 7655.
Cantius ralstoni-USGS 13634; USNM nos. 522157522159, 522168, 540506, 541830-541834; Casts of AMNH 16089 (holotype) and 16092; UW nos. 7063, 7066, 8842.
Cantius torresi-UM nos. 66143, 83470 [holotype], 83475, 87341, 87852, 101958.

## Cantius mckennai-USNM 533559.

Marcgodotius indicus-GU nos. 7, 40, 44, 45, 46, 49, 51, $52,54,227,600,611,703,727,743,744,1534,1536,1538$, 1544, 1554, 1575, 1591, 1602 (Rose et al., 2009).
Asiadapis cambayensis-GU nos. 6, 32, 35-38, 598, $642,745,1505,1537,1627,1649$ (Rose et al., 2009).
Purgatorius janisae-Casts of UCMP 107406 (holotype), 107409; LACM 28128. Other data from Clemens (2004).
Palaechthon-Uppers are scored based on P. nacimienti (UKMNH 9557, cast); lower tooth characters partly from P. nacimienti (UKMNH 9559, holotype, cast). Some lower characters are scored based on P. alticuspis (casts of USNM 9532 [holotype] and AMNH nos. 35479 and 35488) due to availability and the fact that the holotype of P. nacimienti is heavily worn, obscuring some details and affecting some indices. Character 27 is scored as 0 because of elevation of the paraconid, though it may be distinct or less distinct. Characters 21 and 44 are scored as polymorphic to reflect more primitive conditions of P. nacimienti (Silcox, 2001).

## APPENDIX D: RAW DATA FOR $P_{4}$ ANALYSES

| Taxon | Spec. no. | L | $\begin{gathered} \text { Mtd } \\ \text { h1 } \end{gathered}$ | $\begin{gathered} \text { Prt } \\ \text { h } \end{gathered}$ | $\begin{gathered} \text { Mtd } \\ \text { h2 } \end{gathered}$ | $\begin{aligned} & \text { Tri } \\ & \text { W } \end{aligned}$ | $\begin{gathered} \text { Tri } \\ \text { L } \end{gathered}$ | $\begin{aligned} & \mathrm{Pp} \\ & \mathrm{Vt} \\ & \mathrm{Ag} \end{aligned}$ | $\begin{gathered} \mathrm{Bl} \\ \mathrm{Ag} \end{gathered}$ | $\begin{gathered} \mathrm{Mes} \\ \mathrm{Ag} \end{gathered}$ | $\begin{aligned} & \text { Tri } \\ & \text { W } \\ & \text { In } \end{aligned}$ | $\begin{gathered} \text { Mtd } \\ \text { h1 } \\ \text { In } \\ \hline \end{gathered}$ | $\begin{gathered} \text { Mtd } \\ \text { h2 } \\ \text { In } \\ \hline \end{gathered}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Teil. asiatica | IVPP 7165 | 1.43 | 0.84 | 1.35 | 0.79 | 1.20 | 0.87 | 42 | 36 | 142 | 1.37 | 0.63 | 0.548 |
| Teil. magnoliana? | CM 70434 | 1.39 | 0.79 | 1.06 | 0.69 | 1.17 | 0.68 | 53 | 54 | 119 | 1.73 | 0.74 | 0.500 |
| Teil. magnoliana | CM 73229 | br | 0.98 | 1.40 | 0.83 | 1.19 | 0.95 | 48 | 40 | 135 | 1.26 | 0.70 | - |
| Teil. belgica | IRSNB CL 192 | 1.33 | 0.88 | 1.36 | 0.77 | 1.05 | 0.86 | 48 | 40 | 127 | 1.23 | 0.65 | 0.575 |
| Teil. belgica | IRSNB CL 246 | 1.53 | 0.90 | 1.37 | 0.76 | 1.16 | 0.92 | 51 | 44 | 124 | 1.26 | 0.66 | 0.497 |
| Teil. belgica | IRSNB CL 457 | 1.42 | 0.89 | 1.35 | 0.75 | 1.10 | 0.87 | 48 | 41 | 123 | 1.26 | 0.66 | 0.532 |
| Teil. belgica | IRSNB M64 | 1.38 | 0.91 | 1.40 | 0.77 | 1.08 | 0.78 | 49 | 38 | 139 | 1.38 | 0.65 | 0.556 |
| Teil. belgica | IRSNB M4090 | 1.48 | 0.95 | 1.39 | 0.79 | 1.21 | 0.89 | 50 | 42 | 129 | 1.36 | 0.68 | 0.531 |
| Teil. belgica | IRSNB M4291 | br | 0.86 | 1.32 | 0.73 | br | br | br | 41 | br | - | 0.66 | - |
| Teil. belgica | IRSNB M4292 | 1.44 | 0.85 | 1.33 | 0.69 | 1.07 | 0.85 | 51 | 43 | 123 | 1.25 | 0.64 | 0.479 |
| Teil. belgica | IRSNB WL 13 | 1.41 | 0.83 | 1.26 | 0.76 | 1.09 | 0.93 | 51 | 40 | 128 | 1.17 | 0.66 | 0.539 |
| Teil. belgica | IRSNB WL 159 | 1.35 | 0.78 | 1.26 | 0.66 | 1.11 | 0.87 | 43 | 40 | 141 | 1.27 | 0.62 | 0.491 |
| Teil. belgica | IRSNB WL 1052 | 1.49 | 0.92 | 1.34 | 0.84 | 1.14 | 0.86 | 47 | 44 | 126 | 1.33 | 0.69 | 0.564 |
| Teil. belgica | IRSNB WL 1062 | 1.31 | 0.87 | 1.20 | 0.81 | 0.99 | 0.77 | 52 | 42 | 125 | 1.29 | 0.73 | 0.618 |
| Teil. belgica | IRSNB WL 1180 | 1.37 | 0.91 | 1.28 | 0.84 | 1.05 | 0.84 | 47 | 44 | 129 | 1.25 | 0.71 | 0.613 |
| Teil. brandti | UF 244454 | 1.49 | 0.95 | 1.37 | 0.93 | 1.34 | 1.05 | 46 | 44 | 126 | 1.28 | 0.69 | 0.625 |
| Teil. brandti | USNM 493913 | 1.48 | 0.87 | 1.44 | 0.88 | 1.22 | 0.93 | 44 | 45 | 129 | 1.31 | 0.60 | 0.591 |
| Teil. brandti | USNM 525543 | 1.56 | 0.94 | 1.31 | 0.95 | 1.29 | 0.91 | 51 | 45 | 125 | 1.43 | 0.72 | 0.608 |
| Teil. brandti | USNM 525544 | 1.57 | 1.02 | 1.46 | 0.90 | 1.28 | 0.97 | 44 | 50 | 133 | 1.32 | 0.70 | 0.577 |
| Teil. brandti | USNM 533554 | 1.59 | 0.97 | 1.36 | 0.90 | 1.24 | 0.95 | 48 | 41 | 136 | 1.30 | 0.72 | 0.564 |
| Teil. americana | UF 244452 | 1.36 | 0.94 | 1.26 | 0.87 | 1.19 | 0.88 | 59 | 55 | 124 | 1.35 | 0.75 | 0.638 |
| Teil. americana | UM 65770 | 1.54 | 1.01 | 1.42 | 0.75 | 1.25 | 0.98 | 61 | 49 | 122 | 1.28 | 0.71 | 0.486 |
| Teil. americana | UM 75160 | 1.37 | 1.05 | 1.35 | 0.94 | 1.36 | 0.97 | 61 | 50 | 131 | 1.41 | 0.78 | 0.690 |
| Teil. americana | UM 76600 | 1.47 | 1.05 | 1.44 | 0.87 | 1.36 | 0.92 | 60 | 53 | 128 | 1.47 | 0.73 | 0.592 |
| Teil. americana | USGS 3849 | 1.47 | 0.96 | 1.19 | 0.98 | 1.21 | 1.02 | 72 | 53 | 127 | 1.19 | 0.81 | 0.667 |
| Teil. americana | USGS 15406 | 1.49 | 1.11 | 1.47 | 0.94 | 1.45 | 1.08 | 62 | 52 | 127 | 1.34 | 0.76 | 0.631 |
| Teil. americana | USGS 25324 | 1.35 | 0.87 | 1.21 | 0.73 | 1.21 | 0.92 | 56 | 47 | 142 | 1.31 | 0.72 | 0.541 |
| Teil. americana | USNM 539489 | 1.47 | 0.95 | 1.28 | 0.89 | 1.27 | 0.91 | 76 | 48 | 118 | 1.40 | 0.74 | 0.605 |
| Teil. americana | UW 7165 | 1.39 | 1.00 | 1.38 | 0.85 | 1.38 | 0.95 | 57 | 51 | 127 | 1.46 | 0.73 | 0.608 |
| Teil. crassidens | UM 71071 | 1.32 | 1.03 | 1.21 | 0.92 | 1.34 | 0.91 | 63 | 69 | 116 | 1.47 | 0.85 | 0.697 |
| Teil. crassidens | UM 71126 | 1.37 | 1.09 | 1.30 | 1.02 | 1.32 | 0.95 | 66 | 63 | 110 | 1.39 | 0.84 | 0.745 |
| Teil. crassidens | UM 71386 | 1.35 | 1.09 | 1.22 | 0.99 | 1.28 | 0.90 | 64 | 57 | 116 | 1.42 | 0.89 | 0.733 |
| Marcgodinotius indicus | GU 40 | na | nm | na | nm | 1.31 | 1.40 | 42 | 38 | 139 | 0.94 | - | - |
| Marcgodinotius indicus | GU 727 | 1.72 | 0.97 | 1.71 | 0.45 | 1.35 | 1.41 | 44 | 36 | 126 | 0.96 | 0.57 | 0.262 |
| Marcgodinotius indicus | GU 1536 | na | nm | na | nm | 1.34 | 1.40 | 38 | 35 | 140 | 0.96 | - | - |
| Asiadapis cambayensis | GU 38 | na | nm | na | nm | 1.69 | 1.85 | 38 | 37 | 138 | 0.91 | - | - |
| Asiadapis cambayensis | GU 1627 | na | nm | na | nm | 1.59 | 1.57 | 34 | 38 | 136 | 1.01 | - | ${ }^{-}$ |
| Altanius orlovi | PSS 20-58 | na | na | na | na | na | na | 41 | 40 | 151 | 1.15 | 0.83 | 0.760 |
| Tetonius homunculus | AMNH 4194-1 | na | na | na | na | na | na | 78 | 56 | 112 | 1.55 | 0.77 | 0.584 |
| Tetonius matthewi | USGS 3841 | 2.07 | 1.35 | 1.73 | 1.11 | 1.97 | 1.37 | 71 | 56 | 118 | 1.44 | 0.78 | 0.536 |
| Tetonius matthewi | USNM 487864 | 1.72 | 1.16 | 1.55 | 1.03 | 1.53 | 1.08 | 63 | 52 | 124 | 1.42 | 0.75 | 0.599 |
| Tetonius matthewi | USNM 487865 | 1.94 | 1.25 | 1.62 | 1.15 | 1.72 | 1.23 | 69 | 57 | 113 | 1.40 | 0.77 | 0.593 |
| Tetonius matthewi | USNM 487870 | 1.98 | 1.35 | 1.85 | 1.25 | 1.95 | 1.42 | 64 | 53 | 122 | 1.37 | 0.73 | 0.631 |
| Tetonius matthewi | USNM 527713 | 1.90 | 1.32 | 1.97 | 1.17 | 1.92 | 1.32 | 63 | 51 | 121 | 1.45 | 0.67 | 0.616 |
| Tetonius matthewi | USNM 533455 | 2.02 | 1.49 | 2.09 | 1.39 | 1.99 | 1.41 | 66 | 56 | 120 | 1.41 | 0.71 | 0.688 |
| Tetonius matthewi | YPM 23031 | 1.82 | 1.30 | 1.75 | 1.19 | 1.79 | 1.30 | 65 | 54 | 116 | 1.38 | 0.74 | 0.654 |
| Donrussellia gallica | composite | na | na | na | na | na | na | 37 | 33 | 153 | 0.93 | 0.59 | 0.483 |
| Steinius vespertinus | USGS 25027 | na | na | na | na | na | na | 56 | 49 | 125 | 1.40 | 0.74 | 0.668 |
| Purgatorius janisae | UCMP 107406 | na | na | na | nm | na | na | 34 | 29 | 143 | 1.10 | - | - |
| Palaechthon alticuspis | AMNH 35488 | na | na | na | na | na | na | 59 | 26 | 138 | 0.96 | 0.75 | 0.687 |
| Cantius ralstoni | UW 8842 | na | na | na | na | na | na | 36 | 39 | 144 | 1.10 | 0.77 | 0.590 |

Abbreviations: Ag, angle; Bl, buccolingual; br, broken morphology; h, height; In, index; L, length; Mtd, metaconid; Mes, Mesial; na, measurement not available; nm, necessary morphology not present; Pp, preprotocristid; Prt, protoconid; Tri, trigonid; W, width; Vt, verticality. Angles measured in degrees; linear measurements in mm. Metaconid height 1 refers to character 28; metaconid height 2 is character 47.
APPENDIX E: $P_{4}$ AND $M_{2}$ CHARACTERS AND STATISTICS: (A) $P_{4}$ AND $M_{2}$ CHARACTERS, DESCRIPTIVE STATISTICS, AND CODINGS

|  | Pj | Pan | Ao | Cr | Dp | Dg | Mi | Asc | Tas | Tbg | Tbr | Tmg | Ta | Tc | Ttm | Sto |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Character 26: $\mathrm{P}_{4}$ buccolingual angle ( $0,<39 ; 1,39-55 ; 2,>55$ ) |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| s.d. | 29, na | 26, na | 40, na | 39, na | ? | 33, na | 36.33, 1.5 | 37.5, na | 36, na | 41.6, 2.0 | 44.9, 3.3 | 40, na | 50.9, 2.6 | 63.1, 5.9 | 54, 2.3 | 49, na |
| range | na | na | na | na | ? | na | 35-38 | 37-38 | na | 38-44 | 41-50 | na | 47-55 | 57-69 | 51-57 | Na |
| code | 0 | 0 | 1 | 1 | ? | 0 | 0 | 0 | 0 | 1 | 1 | 1 | 1 | 2 | 2 | 1 |
| Character 28: $\mathrm{P}_{4}$ metaconid height $1(0,<0.60 ; 1,0.60-0.68 ; 2,0.69-0.78 ; 3,>0.78)$ |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| x , s.d. | na, na | 0.75, na | 0.83, na | 0.77, na | ? | 0.59 , na | 0.57, na | na, na | 0.63, na | 0.67, 0.03 | 0.68, 0.2 | 0.72, na | 0.75, 0.03 | 0.86, 0.02 | 0.73, 0.03 | 0.74, na |
| range | na | na | na | na | ? | na | na | na | na | 0.62-0.73 | 0.60-0.72 | 0.70-0.74 | 0.71-0.81 | 0.84-0.89 | 0.67-0.77 |  |
| code | ? | 2 | 3 | 2 | ? | 0 | 0 | na | 1 | 1 | 1,2 | 2 | 2 | 3 | 2 | 2 |
| Character 29: $\mathrm{P}_{4}$ preprotocristid verticality angle ( $0,<43 ; 1,43-52 ; 2,53-62 ; 3,>63$ ) |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| x , s.d. | 34, na | 59, na | 41, na | 36, na | ? | 37, na | 41.33,3.0 | 36, na | 42 , na | 48.8, 2.6 | 46.6, 3.0 | 48, na | 63, 6.8 | 64.33,1.53 | 66, 3.07 | 56, na |
| range | na | na | na | na | ? | na | 38-44 | 34-38 | na | 43-52 | 44-51 | na | 56-76 | 63-66 | 63-71 | na |
| code | 0 | 2 | 0 | 0 | ? | 0 | 0,1 | 0 | 0 | 1 | 1 | 1 | 2 | 3 | 3 | 2 |
| Character 31: $\mathrm{P}_{4}$ trigonid width ( $0,<1.21 ; 1,1.21$ and greater) |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| x , s.d. | 1.1, na | 0.96, na | 1.15, na | 1.1, na | ? | 0.93, na | 0.95, 0.01 | 0.96, na | 1.37, na | 1.29, 0.06 | 1.28, 0.06 | 1.26, na | 1.35, 0.09 | 1.43, 0.04 | 1.43,0.03 | 1.4, na |
| range | na | na | na | na | ? | na | 0.94-0.96 | 0.91-1.01 | na | 1.23-1.38 | 1.17-1.38 | na | 1.19-1.47 | 1.39-1.47 | 1.37-1.55 | na |
| code | 0 | 0 | 0 | 0 | ? | 0 | 0 | 0 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| Character 39: $\mathrm{M}_{2}$ relief ( $0,>0.55 ; 1,0.55-0.53 ; 2,<0.53$ ) |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| x , s.d. | 0.63, na | 0.56, na | 0.55, na | 0.49, 0.02 | 0.5 | 0.53,0.02 | 0.55 , na | 0.52 , na | 0.55, na | 0.50, 0.01 | 0.49, 0.02 | 0.51, na | 0.51, 0.02 | 0.48, 0.01 | ? | ? |
| range | na | na | na | 0.47-0.53 | na | 0.50-0.56 | na | na | na | 0.48-0.52 | 0.47-0.51 | na | 0.48-0.52 | 0.48-0.49 | ? | ? |
| code | 0 | 0 | 1 | 2 | 2 | 2 | 1 | 2 | 1 | 2 | 2 | 2 | 2 | 2 | ? | ? |
| Character 46: $\mathrm{P}_{4}$ mesial angle ( $0,125-145 ; 1,>145 ; 2,<125$ ) |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| x , s.d. | 143, na | 138, na | 151, na | 144, na | ? | 153, na | 135, 7.8 | 137, na | 142, na | 128.5, 6.1 | 129.8, 4.7 | 135, na | 127.3, 6.7 | 114, 3.5 | 119, 3.8 | 125, na |
| range | na | na | na | na | ? | na | 126-140 | 136-138 | na | 123-141 | 125-136 | na | 118-142 | 110-116 | 113-124 | na |
| code | 0 | 0 | 1 | 0 | ? | 1 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 2 | 2 | 0 |
| Character 47: $\mathrm{P}_{4}$ metaconid height $2(0,<0.56 ; 1,0.56-0.66 ; 2,>0.66)$ |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| x, s.d. | na, na | 0.69, na | 0.76, na | 0.59, na | ? | 0.48 , na | 0.26 , na | na | 0.55 , na | 0.54, 0.05 | 0.59, 0.02 | ? | 0.61, 0.06 | 0.72,0.02 | 0.65, 0.05 | 0.67, na |
| range | na | na | na | na | ? | na | na | na | na | 0.48-0.62 | 0.56-0.62 | ? | 0.49-0.69 | 0.70-0.73 | 0.60-0.72 | na |
| code | na | 2 | 2 | 1 | ? | 0 | 0 | na | 0 | 0 | 1 | ? | 0,1 | 2 | 1 | 2 |

x, mean; s.d., standard deviation; na, not applicable/available; Ao, Altanius orlovi; Asc, Asiadapis cambayensis; Cr, Cantius ralstoni; Dg, Donrussellia gallica; Dp, D. provincialis; Mi, Marcgodinotius indicus; Pan, Palaechthon nacimiento; Pj, Purgatorius janisae; Stv, Steinius vespertinus; Ta, Teilhardina americana; Tas, T. asiatica; Tbg, T belgica; Tbr, T. brandti; Tc, T. crassidens; Tmg, T. magnoliana; Ttm, Tetonius matthewi.
APPENDIX E: $P_{4}$ AND $M_{2}$ CHARACTERS AND STATISTICS: (B) RESULTS OF KRUSKAL-WALLIS TEST OF P 4 AND M ${ }_{2}$ CHARACTERS

| Characters | $H$ | $P$ | Tbg v. Tbr | Tbg v. Ta | Tbg v. Tc | Tbr v. Ta | Tbr v. Tc | Ta v. Tc |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $\mathrm{P}_{4} \mathrm{~B}-\mathrm{L}$ angle | 21.9 | $>0.0001$ | 0.027/0.16 | 0.0001/0.001 | 0.013/0.08 | 0.016/0.1 | 0.037/0.22 | 0.016/0.1 |
| $\mathrm{P}_{4}$ metaconid height 1 | 20.2 | 0.0002 | ns | 0.0003/0.002 | 0.011/0.07 | 0.011/0.07 | 0.036/0.22 | 0.016/0.1 |
| $\mathrm{P}_{4}$ preprot verticality | 21.0 | $>0.0001$ | ns | 0.0002/0.001 | 0.013/0.08 | 0.003/0.02 | 0.036/0.22 | ns |
| $\mathrm{P}_{4}$ trigonid width | 10.3 | 0.016 | ns | 0.03/0.2 | 0.013/0.08 | ns | ns | ns |
| $\mathrm{M}_{2}$ relief | 6.1 | 0.106 | ns | ns | ns | ns | ns | ns |
| $\mathrm{P}_{4}$ mesial angle | 8.5 | 0.038 | ns | ns | 0.012/0.07 | ns | 0.036/0.22 | 0.015/0.09 |
| $\mathrm{P}_{4}$ metaconid height 2 | 13.03 | 0.005 | ns | 0.03/0.2 | 0.013/0.08 | ns | 0.037/0.22 | 0.016/0.1 |


 ferroni-corrected. "ns" is used for comparisons that were non-significant prior to correcting for multiple comparisons.


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[^1]:    ${ }^{1}$ Characters have been modified after Seiffert et al. (2005), Smith et al. (2006), and Beard (2008) to reflect nuances in character states in the principal taxa and to clarify ambiguities or quantify characters deemed otherwise too subjective to be repeatable. All taxa were reexamined based on available material (original specimens or casts) or literature. These new observations resulted in revision of the scores for several characters. Nearly all such cases pertain to subjective characters which are closer to the boundary between character states than to the extremes. Even when characters were quantified, ambiguity remains because quantitative boundaries for individual character states are necessarily arbitrary. Character scores were based on means of the available specimens or measurements, but ranges for taxa may encompass more than one character state. When only two or three specimens were available, characters were scored as polymorphic if the specimens differed substantially. Moreover, it should be obvious that a slight shift in the arbitrary boundaries may alter the character scores, which could easily affect the outcome of the analysis. Wherever possible, boundaries were imposed in this analysis where natural gaps occur.

